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# New Aromadendrane Sesquiterpenoid Pseuboydone F from the Marine-derived Fungus *Pseudallescheria boydii* F44-1

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**Abstract:** A new aromadendrane sesquiterpenoid pseuboydone F (1), along with a known pseuboydone A (2), were isolated from the marine-derived fungus *Pseudallescheria boydii* F44-1 associated with the soft coral *Sarcophyton* sp.. The structures were elucidated by HRMS, 1D and 2D NMR spectroscopic data.

**Keywords:** Marine fungus; *Pseudallescheria boydii*; aromadendrane; sesquiterpenoid; pseudboydone F. © 2019 ACG Publications. All rights reserved.

## **1. Introduction**

Marine-derived fungi are considered as a promising source of novel natural products with biodiversity and chemical diversity. Although numerous novel metabolites have been obtained from a large number of marine-derived fungi, actually the discovering rate is still not high. It's challengeable to develop efficient dereplication techniques based on NMR or MS analysis [1,2]. In our previous studies, we reported the strategies to increase the discovery rate of new compounds by tracking the diagnostic <sup>1</sup>H or <sup>13</sup>C NMR resonance signals. For example, a pair of unprecedented epimonothiodiketopiperazine diastereomers, pseudellones A and B were isolated from the marine fungus *Pseudallescheria ellipsoidea* by tracking the relatively rare <sup>1</sup>H NMR resonated signals in the range of 8.00-8.50 ppm [3]. Following with the rich aromatic proton signals in aromatic range of 6.5-8.5 ppm, 14 new alkaloids were obtained from marine-derived fungus *Scedosporium apiospermum* F41-1 [4].

Recently, we have isolated a fungal strain *Pseudallescheria boydii* (collection No. F44-1) from the soft coral *Sarcophyton* sp. collected in the Hainan Sanya National Coral Reef Reserve, China. This fungus was cultured in glucose-peptone-yeast extract (GPY) media and prescreened the metabolites extract using the <sup>1</sup>H NMR spectroscopy. At the high field area, two sets of signals at  $\delta_{\rm H}$  0.50 (dd, 9.6,

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9.6) and  $\delta_{\rm H}$  0.17 (dd, 9.6, 9.6) attracted our attention. To the best of our knowledge, the aromadendrane sesquiterpenoids showed the signals in the range of 0-0.6 ppm due to the shielding effect of the cyclopropane ring. Previously, by tracking the proton resonance signals in this region, the aromadendrane sesquiterpenoids pseuboydones A and B were separated from marine-derived fungus *Pseudallescheria boydii* F19-1[5], and scedogiines A-F were isolated from the marine-derived fungus *Scedosporium dehoogii* F41-4 [6]. Using the same strategy, a new aromadendrane sesquiterpenoid pseuboydone F (1) and a known pseuboydone A (2) (Figure 1) were isolated from the fungus *Pseudallescheria boydii* F44-1. Herein we report the isolation and structural elucidation of these compounds.

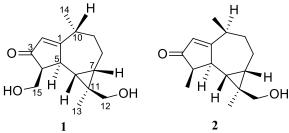


Figure 1. The chemical structures of compounds 1 and 2

## 2. Materials and Methods

#### 2.1. General Experimental Procedures

Preparative HPLC was performed using a Shimadzu LC-20AT HPLC pump (Shimadzu Corporation, Nakagyo-ku, Kyoto, Japan) and installed with an SPD-20A dual  $\lambda$  absorbance detector (Shimadzu Corporation, Nakagyo-ku, Kyoto, Japan) and a Capcell-Pak C18 UG80 HPLC column (250 mm  $\times$  20 mm, Shiseido, Japan). 1D and 2D NMR experiments were measured with Bruker Avance 400 spectrometers and Bruker Avance 600 spectrometers. The chemical shifts are relative to the residual solvent signal (CDCl<sub>3</sub>:  $\delta_{\rm H}$  7.26 and  $\delta_{\rm C}$  77.0). The HR-APCI-MS spectrum was measured with Thermo Obitrap Fusion Lumos liquid chromatography-mass spectrometry.

#### 2.2. Fungal Identification and Culture Method

The marine fungus *Pseudallescheria boydii* (collection number F44-1) was isolated from the inner tissue of the soft coral *Sarcophyton* sp. collected from Hainan Sanya National Coral Reef Reserve, P. R. China. This fungal strain was conserved in 15% (v/v) glycerol aqueous solution at -80 °C. A voucher specimen was deposited in the School of Chemistry, Sun Yat-sen University, Guangzhou, P. R. China. Analysis of the ITS rDNA by BLAST database screening provided 99.9% match to *Pseudallescheria boydii*. The marine fungus *Pseudallescheria boydii* F44-1, was cultured in the GPY medium which included 15 g/ L glucose, 5 g/ L peptone, 2 g/ L yeast extract, 25 g/ L sea salt, and 1 L H<sub>2</sub>O at pH 7.0. Fungal mycelia were cut and transferred aseptically to 1000 mL conical flasks each containing 600 mL sterilized liquid medium. The fungus was incubated at 28 °C for 20 days.

#### 2.3. Extraction and Isolation

10 liters culture broth was filtered through cheesecloth. The liquid was successively extracted three times with EtOAc ( $3\times10$  L). Finally, the extract was concentrated by low-temperature rotary evaporation to get a crude extract (2.8 g). The extract was chromatographed on a silica gel column (diameter: 4 cm, length: 50 cm, silica gel, 35 g) with a gradient of petroleum Ether-EtOAc (30:0-0:30, v/v) followed by EtOAc-MeOH (30:0-0:30, v/v) to yield ten fractions (Fr.1-Fr.10). The fractions were monitored by TLC and similar fractions, Fr.6-Fr.8 were combined and concentrated in vacuo, and then, the constituents was purified by silica gel column using a step gradient elution with ether-EtOAc (10:0-0:10, v/v) to get 10 subfractions (Fr.6-8-1 to Fr.6-8-10). Then Fr.6-8-5 was further purified using reversed phase preparative HPLC with a mobile phase of CH<sub>3</sub>CN-H<sub>2</sub>O (60:40, v/v, t<sub>R</sub> = 30 min)

to obtain compound **1** (1.2 mg). Further HPLC purification of Fr.6-8-6 with CH<sub>3</sub>OH-H<sub>2</sub>O (75:25, v/v,  $t_R = 37.5$  min) gave compound **2** (3.9 mg).

## 3. Results and Discussion

#### 3.1. Structure Elucidation

Pseudboydone F (1) was obtained as a pale yellow oil. The molecular formula was determined to be  $C_{15}H_{22}O_3$  by the HR-APCI-MS peak at m/z 249.14906 [M-H]<sup>-</sup> (calcd  $C_{15}H_{21}O_3$ , 249.14962) indicating five degrees of unsaturation. The <sup>13</sup>C NMR and DEPT spectra displayed two sp<sup>2</sup> quaternary carbons, one sp<sup>3</sup> quaternary carbon, one sp<sup>2</sup> methine, five sp<sup>3</sup> methines, four sp<sup>3</sup> methylenes and two methyls (Table 1). The <sup>1</sup>H-<sup>1</sup>H COSY correlations of H<sub>2</sub>-15/H-4/H-5/H-6/H-7/H<sub>2</sub>-8/H<sub>2</sub>-9/H-10/H<sub>3</sub>-14 revealed the fragment -CH2-CH-CH-CH-CH2-CH2-CH2-CH2-CH3 (Figure 2). In the <sup>1</sup>H NMR spectrum, two characteristic signals at  $\delta_{\rm H}$  0.50 (dd, 9.6, 9.6, H-6) and 0.87 (ddd, 11.4, 9.6, 6.2, H-7) (Table 1) displayed the existence of a cyclopropane ring. One carbonyl group ( $\delta_c$  210.7, C-3) and a trisubstituted double bond (\delta<sub>C</sub> 190.3, C-1; 126.6, C-2) illustrated two degrees of unsaturation. So, compound 1 had to contain another two rings. Further analysis of the HMBC correlations from H-2 to C-4 and C-5, from H<sub>3</sub>-14 to C-1, the olefinic quaternary carbon C-1 was connected to C-5 ( $\delta_{C}$  44.0) forming a bridge. A five-membered ring system was constructed by quaternary carbon C-1 and methine C-4 ( $\delta_{\rm C}$  57.9) via carbonyl group C-3. Therefore, compound 1 contains a five- and sevenmembered rings and cyclopropane fused ring system, which belong to aromadendrane sesquiterpenoid. In addition, one oxygenated methylene and one methyl connected with C-11 ( $\delta_{C}$ 27.4), which was confirmed by the HMBC correlations from H<sub>2</sub>-12 to C-6, C-7 and C-11, from H<sub>3</sub>-13 to C-11 and C-12. The remaining one oxygenated methylene C-15 was attached to the C-4 position based on the HMBC correlations from H<sub>2</sub>-15 to C-4 and C-5.

No.	1ª		2 <sup>b</sup>	
	$\delta_{ m C}$	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m H}$
1	190.3, C		187.7, C	
2	126.6, CH	5.86 (d, 1.2)	125.2, CH	5.81 (s)
3	210.7, C		211.5, C	
4	57.9, CH	2.50 (dd, 6.6, 6.6)	46.7, CH	2.60 (m)
5	44.0, CH	2.45 (d, 9.6)	44.3, CH	2.60 (m)
6	28.5, CH	0.50 (dd, 9.6, 9.6)	25.2, CH	0.17 (dd, 9.6, 9.6)
7	25.7, CH	0.87 (ddd, 11.4, 9.6, 6.2)	25.0, CH	0.83 (m)
8	23.5, CH <sub>2</sub>	1.27 (m)	23.2, CH	1.24 (m)
		2.07 (ddd, 14.4, 6.6, 6.2)		2.06 (m)
9	35.1, CH <sub>2</sub>	1.44 (ddd, 12.6, 12.6, 12.0)	35.8, CH <sub>2</sub>	1.36 (m)
		1.97 (ddd, 12.6, 6.6, 6.6)		2.00 (m)
10	40.7, CH	2.35 (ddq, 12.6, 6.6, 6.6)	40.2, CH	2.34 (m)
11	27.4, C		26.7, C	
12	72.6, CH <sub>2</sub>	3.29 (d, 10.8)	72.3, CH <sub>2</sub>	3.24 (d, 10.8)
		3.39 (d, 10.8)		3.43 (d, 10.8)
13	11.4 CH <sub>3</sub>	1.24 (s)	11.5, CH <sub>3</sub>	1.20 (s)
14	20.1, CH <sub>3</sub>	1.26 (d, 6.6)	19.7, CH <sub>3</sub>	1.24 (d, 6.8)
15	63.4, CH <sub>2</sub>	3.69 (dd, 10.2, 6.6)	10.0, CH <sub>3</sub>	1.13 (d, 6.4)
		3.76 (dd, 10.2, 6.6)		
				1 1

**Table 1**. <sup>1</sup>H and <sup>13</sup>C NMR data of compounds **1** and **2** in CDCl<sub>3</sub> ( $\delta$  in ppm, *J* in Hz).

<sup>a 13</sup>C NMR data were recorded at 150 MHz and <sup>1</sup>H NMR data were recorded at 600 MHz.

<sup>b 13</sup>C NMR data were recorded at 100 MHz and <sup>1</sup>H NMR data were recorded at 400 MHz.

The relative configuration of compound **1** was established by analysis the NOESY spectrum. The cross peaks of H-6/H-7, H-6/H<sub>2</sub>-12, H-6/  $H_2$ -15, and H-7/H<sub>2</sub>-12, implied that H-6, H-7, H<sub>2</sub>-12 and

H<sub>2</sub>-15 having the same  $\beta$ -oriented. The NOE interactions of H-5/H<sub>3</sub>-13 and H-5/H<sub>3</sub>-14 suggested H-5, H<sub>3</sub>-13 and H<sub>3</sub>-14 had an  $\alpha$ -orientation.

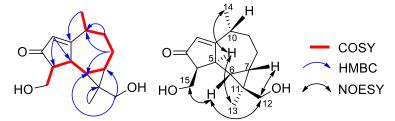


Figure 2. <sup>1</sup>H-<sup>1</sup>H COSY, key HMBC and key NOESY correlations of 1

Followed the triplet at  $\delta_{\rm H}$  0.17 (dd, 9.6, 9.6), compound 2 was purified. By comparing its NMR data with the literature values, compound 2 was identified as pseuboydone A [5].

#### 3.2. Cytotoxicity

Seven cancer cell lines, including CNE1, CNE2, HONE1, SUNE1, A549, GLC82 and HL7702 were used to examine the cytotoxic activities of compounds 1 and 2 *in vitro*. This assay revealed that 1 and 2 were apparently inactive (IC<sub>50</sub> values > 100  $\mu$ M).

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#### **Supporting Information**

Supporting information accompanies this paper on <u>http://www.acgpubs.org/journal/records-of-natural-products</u>

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