

Rec. Nat. Prod. 18:5 (2024) 538-543

records of natural products

# Chemical Composition and Evaluation of the Antibacterial, Synergistic Antibacterial, Antioxidant and Cytotoxic Activities of the Essential Oil of *Macrothelypteris torresiana* (Gaudich.) Ching Xiangyi Li<sup>1</sup>, Shu Qiu<sup>1</sup>, Shiyu Song<sup>1</sup> and Pengxiang Lai<sup>1</sup>, 2

<sup>1</sup> Sdu-Anu Joint Science College, Shandong University, Weihai, 264209, P. R. China <sup>2</sup> Marine College, Shandong University, Weihai 264209, P. R. China

(Received July 25, 2024; Revised September 25, 2024; Accepted October 04, 2024)

**Abstract:** In this study, we evaluated the chemical composition, antioxidant, cytotoxic, and antibacterial activities of the essential oil extracted from the aerial parts of *Macrothelypteris torresiana* (Gaudich.) Ching (MT-EO), as well as its synergistic antibacterial effect in combination with commercial antibiotics. Fifty-seven compounds were identified in MT-EO, representing 97.9% of the total oil content. The compounds bicyclogermacrene (12.9%), spathulenol (11.9%), β-elemene (5.7%), and hexyl hexanoate (5.0%) were detected as the main constituents. The microdilution and checkerboard assays were used to evaluate the antibacterial and synergistic properties of the essential oil. It was found that MT-EO possessed bactericidal activity against all tested bacteria, with MIC values between 0.625 to 1.250 mg/mL, which was the same as MBCs. Additionally, synergistic effects were detected in both *M. torresiana* essential oil -chloramphenicol and -streptomycin combinations. Besides, according to the MTT test, MT-EO possessed broadspectrum cytotoxicities on various cell lines with IC<sub>50</sub> values ranging from 15.12 ± 0.96 to 47.07 ± 1.96 μg/mL, including the MCF-7, A-549, HCT-116, HepG2, and LO2 cell lines. Furthermore, MT-EO showed moderate antioxidant activities in DPPH, ABTS, and FRAP assays, with IC<sub>50</sub> values of 434.5 ± 9.6 and 98.1 ± 1.1 μg/mL, and Trolox equivalent of 97.11 ± 3.37 μmol Trolox × g<sup>-1</sup>, respectively.

**Keywords:** *Macrothelypteris torresiana*; essential oil; antibacterial; synergistic; antioxidant; cytotoxic. © 2024 ACG Publications. All rights reserved.

### 1. Plant Source

The aerial parts of *Macrothelypteris torresiana* (Gaudich.) Ching were collected in August 2022 from Jieyang, Guangdong Province, China. The botanical identification was conducted by Prof. Hong Zhao, Shandong University, China. A herbarium specimen of the plant was stored at the herbarium of the Institute of Botany, Chinese Academy of Sciences (PE 01768725).

# 2. Previous Studies

Macrothelypteris torresiana is a perennial fern of the Thelypteridaceae family, native to the W. Indian Ocean, tropical and subtropical regions of Asia, and Pacific Islands [1]. The aerial part of M. torresiana is used to treat fever, pain, and granulation in Pakistan, India, and China [2]. Additionally, it is employed in traditional Chinese medicine to alleviate edema in individuals suffering from renal disorders [3]. Previous studies have demonstrated the renoprotective potential of the total polyphenols fraction

<sup>\*</sup> Corresponding author: E-Mail: whlhy@sdu.edu.cn

derived from *Macrothelypteris torresiana* through ameliorating oxidative stress and proinflammatory cytokines [3]. Furthermore, research on phytochemistry indicated *M. torresiana* contains various constituents, including flavonoids, terpenoids, and glycosides [4-6], and the potential biological properties have been examined, such as antitumor, hepatoprotective, anti-inflammatory, and antimicrobial activities [7, 8]. However, there have been no reported studies on the essential oil of *M. torresiana*.

# 3. Present Study

The aerial parts of M. torresiana were subjected to hydrodistillation to extract the essential oil. The yield of essential oil was  $0.15 \pm 0.03\%$  (w/w) based on dry weight. The chemical composition of MT-EO was analyzed using GC/FID and GC/MS. As shown in Figure S1 and Table 1, a total of fifty-seven components were identified, which account for 97.9% of the overall MT-EO. Sesquiterpenes are the most abundant chemical class in MT-EO, and the relative amounts were 40.6% in the volatile fractions. The second major chemical class was oxygenated sesquiterpenes with 37.2%, followed by oxygenated monoterpenes with 4.5%. The major compounds were identified as bicyclogermacrene (12.9%), spathulenol (11.9%),  $\beta$ -elemene (5.7%), hexyl hexanoate (5.0%),  $\delta$ -elemene (3.9%), isospathulenol (3.5%), globulol (3.4%), and epi- $\alpha$ -cadinol (3.3%). Spathulenol, one of the major constituents of MT-EO, has been found to possess a broad spectrum of biological activities, including antioxidant, anti-inflammatory, antiproliferative, antimycobacterial, and anti-M. tuberculosis capabilities [9, 10].  $\beta$ -Elemene, known for its anticancer properties against a variety of cell lines, has been demonstrated to possess anti-proliferative effects via triggering apoptosis [11] and antibacterial activity against Mycobacterium tuberculosis strain H37Ra [12]. Besides, bicyclogermacrene exhibited significant cytotoxic potential against HL-60 cells [13].

**Table 1**. The essential oil composition of *M. torresiana* 

| No. | Compounds                    | RI a | RI <sub>lit</sub> b | RI range               | %    | Identificatio<br>n Method |
|-----|------------------------------|------|---------------------|------------------------|------|---------------------------|
| 1   | 1-Octen-3-ol                 | 972  | 974°                | 967-991 <sup>f</sup>   | 0.5  | MS, RI                    |
| 2   | Hexyl acetate                | 1011 | 1007 <sup>c</sup>   | $999-1020^{\rm f}$     | 0.4  | MS, RI                    |
| 3   | 2-Nonanol                    | 1093 | 1097°               | 1076-1120 <sup>f</sup> | 0.3  | MS, RI                    |
| 4   | Linalool                     | 1099 | $1097^{g}$          | 1098-1101 <sup>g</sup> | 0.3  | MS, RI                    |
| 5   | <i>n</i> -Nonanal            | 1103 | 1100°               | 1093-1118 <sup>f</sup> | 1.5  | MS, RI                    |
| 6   | 4-Ethylbenzaldehyde          | 1161 | 1169e               | 1144-1197 <sup>e</sup> | 0.3  | MS, RI                    |
| 7   | $\beta$ -Cyclocitral         | 1219 | 1217 <sup>c</sup>   | 1205-1225 <sup>f</sup> | 0.3  | MS, RI                    |
| 8   | $\delta$ -Elemene            | 1336 | 1335 <sup>c</sup>   | 1327-1344 <sup>f</sup> | 3.9  | MS, RI                    |
| 9   | Cyclosativene                | 1372 | 1369 <sup>c</sup>   | 1360-1380 <sup>f</sup> | 1.4  | MS, RI                    |
| 10  | Hexyl hexanoate              | 1383 | 1382 <sup>c</sup>   | 1371-1399e             | 5.0  | MS, RI                    |
| 11  | isoLongifolene               | 1387 | 1389 <sup>c</sup>   | 1373-1425 <sup>e</sup> | 1.3  | MS, RI                    |
| 12  | $\beta$ -Elemene             | 1391 | 1389 <sup>c</sup>   | 1374-1402 <sup>f</sup> | 5.7  | MS, RI                    |
| 13  | (2E)-Hexenyl caproate        | 1395 | 1385 <sup>c</sup>   | 1385°                  | 0.8  | MS, RI                    |
| 14  | α-Gurjunene                  | 1408 | 1409 <sup>c</sup>   | 1394-1421 <sup>f</sup> | 2.8  | MS, RI                    |
| 15  | Dihydrodehydro-β-ionone      | 1415 | 1424e               | 1424e                  | 1.0  | MS, RI                    |
| 16  | $(E)$ - $\alpha$ -Ionone     | 1426 | 1428 <sup>c</sup>   | 1403-1435 <sup>f</sup> | 0.4  | MS, RI                    |
| 17  | 2-Butyldecahydro-naphthalene | 1429 | 1432 <sup>c</sup>   | 1424-1450 <sup>f</sup> | 0.7  | MS, RI                    |
| 18  | Aromadendrene                | 1438 | 1439 <sup>c</sup>   | 1419-1465 <sup>f</sup> | 1.8  | MS, RI                    |
| 19  | $\alpha$ -Humulene           | 1452 | 1455 <sup>g</sup>   | 1450-1454 <sup>g</sup> | 2.5  | MS, RI                    |
| 20  | allo-Aromadendrene           | 1460 | 1458 <sup>c</sup>   | 1443-1477 <sup>f</sup> | 0.9  | MS, RI                    |
| 21  | Precocene I                  | 1464 | 1461°               | 1461°                  | 0.5  | MS, RI                    |
| 22  | γ-Gurjunene                  | 1471 | 1475°               | 1455-1485 <sup>f</sup> | 1.3  | MS, RI                    |
| 23  | Germacrene D                 | 1480 | 1481 <sup>g</sup>   | 1478-1488 <sup>g</sup> | 0.8  | MS, RI                    |
| 24  | (E)-β-Ionone                 | 1485 | 1487 <sup>c</sup>   | 1470-1498 <sup>f</sup> | 2.4  | MS, RI                    |
| 25  | γ-Amorphene                  | 1491 | 1495 <sup>c</sup>   | 1474-1485 <sup>f</sup> | 0.6  | MS, RI                    |
| 26  | Bicyclogermacrene            | 1495 | 1500°               | 1477-1503 <sup>f</sup> | 12.9 | MS, RI                    |
| 27  | γ-Cadinene                   | 1513 | $1510^{g}$          | 1511-1521 <sup>g</sup> | 2.5  | MS, RI                    |
| 28  | cis-Calamenene               | 1522 | 1528c               | 1511-1541 <sup>f</sup> | 1.6  | MS, RI                    |
| 29  | Elemol                       | 1552 | 1548 <sup>c</sup>   | 1534-1557 <sup>f</sup> | 0.4  | MS, RI                    |
| 30  | (E)-Nerolidol                | 1563 | 1561 <sup>c</sup>   | 1539-1570 <sup>f</sup> | 0.3  | MS, RI                    |

Biological activities of essential oil from Macrothelypteris torresiana

| No. | Compounds                             | RI a | RI <sub>lit</sub> b | RI range               | %    | Identificatio<br>n Method |
|-----|---------------------------------------|------|---------------------|------------------------|------|---------------------------|
| 31  | Palustrol                             | 1566 | 1567°               | 1561-1571 <sup>f</sup> | 0.7  | MS, RI                    |
| 32  | Spathulenol                           | 1578 | 1566 <sup>g</sup>   | 1568-1590 <sup>g</sup> | 11.9 | MS, RI                    |
| 33  | Globulol                              | 1583 | 1590°               | 1568-1592 <sup>f</sup> | 3.4  | MS, RI                    |
| 34  | Viridiflorol                          | 1591 | 1592°               | 1569-1604 <sup>f</sup> | 2.7  | MS, RI                    |
| 35  | Isoaromadendrene epoxide              | 1599 | 1594e               | 1572-1618e             | 0.5  | MS, RI                    |
| 36  | Guaiol                                | 1601 | 1600°               | 1585-1615 <sup>f</sup> | 0.8  | MS, RI                    |
| 37  | Geranyl isovalerate                   | 1609 | 1606 <sup>c</sup>   | 1582-1613 <sup>f</sup> | 1.1  | MS, RI                    |
| 38  | 1,10-di-epi-Cubenol                   | 1614 | 1618 <sup>c</sup>   | 1591-1623 <sup>f</sup> | 1.1  | MS, RI                    |
| 39  | 1-epi-Cubenol                         | 1628 | 1627 <sup>c</sup>   | 1611-1631 <sup>f</sup> | 0.5  | MS, RI                    |
| 40  | Isospathulenol                        | 1638 | $1640^{d}$          | 1621-1641 <sup>f</sup> | 3.5  | MS, RI                    |
| 41  | <i>epi-α</i> -Cadinol                 | 1641 | 1638 <sup>c</sup>   | 1624-1648 <sup>f</sup> | 3.3  | MS, RI                    |
| 42  | <i>epi-α</i> -Muurolol                | 1644 | 1640°               | 1623-1654 <sup>f</sup> | 0.8  | MS, RI                    |
| 43  | α-Cadinol                             | 1654 | 1652 <sup>c</sup>   | 1635-1664 <sup>f</sup> | 0.9  | MS, RI                    |
| 44  | Ylangenol                             | 1663 | 1666 <sup>d</sup>   | 1666 <sup>d</sup>      | 0.8  | MS, RI                    |
| 45  | Elemol acetate                        | 1669 | 1675 <sup>d</sup>   | $1680^{d}$             | 0.7  | MS, RI                    |
| 46  | n-Pentadecanal                        | 1713 | 1715 <sup>d</sup>   | 1703-1728 <sup>f</sup> | 2.1  | MS, RI                    |
| 47  | $\beta$ -Santalol                     | 1717 | 1715 <sup>c</sup>   | 1676-1736 <sup>e</sup> | 1.0  | MS, RI                    |
| 48  | iso-Longifolol                        | 1731 | 1728 <sup>c</sup>   | 1728 <sup>c</sup>      | 0.4  | MS, RI                    |
| 49  | 8α-hydroxy-Eremophila-1,11-dien-9-one | 1780 | $1777^{d}$          | 1777 <sup>d</sup>      | 0.5  | MS, RI                    |
| 50  | Saussurea lactone                     | 1793 | $1806^{d}$          | 1806 <sup>d</sup>      | 0.4  | MS, RI                    |
| 51  | Dehydrosaussurea lactone              | 1831 | $1838^{d}$          | 1838 <sup>d</sup>      | 0.5  | MS, RI                    |
| 52  | Neophytadiene                         | 1836 | 1841 <sup>d</sup>   | 1804-1857e             | 0.3  | MS, RI                    |
| 53  | Hexahydrofarnesyl acetone             | 1843 | $1847^{d}$          | 1831-1855 <sup>f</sup> | 0.8  | MS, RI                    |
| 54  | Valerenic acid                        | 1868 | $1877^{d}$          | 1877 <sup>d</sup>      | 0.5  | MS, RI                    |
| 55  | (E)-2-Hexadecenal                     | 1881 | $1878^{d}$          | 1813-1880 <sup>e</sup> | 1.9  | MS                        |
| 56  | Gazaniolide                           | 1891 | $1894^{d}$          | -                      | 0.7  | MS                        |
| 57  | Hexadecanoic acid                     | 1961 | 1959 <sup>c</sup>   | 1939-1996 <sup>f</sup> | 1.0  | MS, RI                    |
|     | Oxygenated monoterpenes               |      |                     |                        | 4.5  |                           |
|     | Sesquiterpene hydrocarbons            |      |                     |                        | 40.6 |                           |
|     | Oxygenated sesquiterpenes             |      |                     |                        | 37.2 |                           |
|     | Total identification                  |      |                     |                        | 97.9 |                           |

<sup>a</sup>Retention index calculated from n-alkanes ( $C_7$ - $C_{30}$ ) on HP-5MS column; <sup>b</sup>Linear retention indices from literature: <sup>c</sup>[14], <sup>d</sup>[15], <sup>e</sup>[16], <sup>f</sup>[17], <sup>g</sup>[18].

The MT-EO was evaluated for possible antibacterial activity against selected Gram-positive and Gram-negative pathogenic bacterial strains by using a broth microdilution assay [19]. The positive control utilized was chloramphenicol.

**Table 2**. Antibacterial activity of MT-EO

| De stantal stantas                | MIC, μg/n | nL   | MBC, μg/ | MBC, μg/mL |  |
|-----------------------------------|-----------|------|----------|------------|--|
| Bacterial strains                 | MT-EO     | Ch   | MT-EO    | Ch         |  |
| Gram-positive                     |           |      |          |            |  |
| Bacillus subtilis ATCC 6633       | 625.0     | 2.5  | 625.0    | 20.0       |  |
| Staphylococcus aureus ATCC 6538   | 625.0     | 5.0  | 625.0    | 40.0       |  |
| Gram-negative                     |           |      |          |            |  |
| Escherichia coli ATCC 25922       | 625.0     | 2.5  | 625.0    | 40.0       |  |
| Pseudomonas aeruginosa ATCC 27853 | 1250.0    | 40.0 | 1250.0   | 156.3      |  |

Table 2 shows the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) values of MT-EO against the strains tested. As presented in Table 2, MT-EO exhibits broad-spectrum antibacterial activity against all bacteria strains tested, with MIC values ranging from 0.625 to 1.250 mg/ml, which were the same as MBCs, indicating the potent bactericidal activity of MT-EO. MT-EO exhibited weaker antibacterial activities compared to the synthetic antibiotic Chloramphenicol. The main constituents of MT-EO are sesquiterpenoids and oxygenated sesquiterpenes, which are known to have remarkable antibacterial properties and are considered to be responsible for antibacterial activity [20].

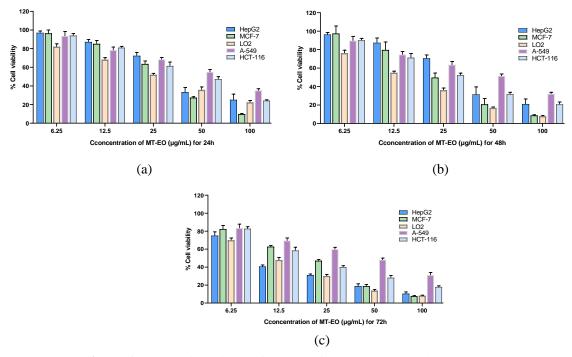
The interaction between MT-EO and conventional antibiotics (chloramphenicol and streptomycin) was assessed using the checkerboard microdilution method [21] based on determined MIC values. The results presented in Table 3 revealed the remarkable synergistic effects of MT-EO combined with antibiotics on all tested bacterial strains, with FICI ranging from 0.15 to 0.37. It is noteworthy that the MIC<sub>c</sub> values of chloramphenicol and streptomycin were observed to be 4-32 and 6-12 times lower than those normally required for direct inhibition of bacterial growth. The results suggested that combining MT-EO with traditional synthetic antibiotics could be an effective way to enhance antibiotic antimicrobial efficacy, expanding the range of antimicrobial activity, preventing resistance, and reducing harmful or undesired side effects [22].

| <b>Table 3.</b> Effect of combination of M' | O with Chloram | phenicol and Streptomycin |
|---|----------------|---------------------------|
|---|----------------|---------------------------|

| Strains                  | Sample | MICa   | MICc  | FICI     | sample | MICa   | MICc  | FICI     |
|--------------------------|--------|--------|-------|----------|--------|--------|-------|----------|
| B. subtilis ATCC 6633    | MT-EO  | 625.0  | 156.2 | 0.37 (S) | MT-EO  | 625.0  | 39.1  | 0.22 (8) |
| B. subillis ATCC 0033    | Ch     | 2.4    | 0.3   | 0.57 (3) | Sm     | 0.6    | 0.1   | 0.23 (S) |
| S. aureus ATCC 6538      | MT-EO  | 625.0  | 39.1  | 0.31 (S) | MT-EO  | 625.0  | 78.1  | 0.21 (8) |
| s. dureus ATCC 0538      | Ch     | 4.9    | 1.2   | 0.51 (5) | Sm     | 1.2    | 0.1   | 0.21 (S) |
| E. coli ATCC 25922       | MT-EO  | 625.0  | 78.1  | 0.25 (S) | MT-EO  | 625.0  | 78.1  | 0.25 (8) |
| E. Coll ATCC 23922       | Ch     | 2.4    | 0.3   |          | Sm     | 2.4    | 0.3   | 0.25(S)  |
| P. aeruginosa ATCC 27853 | MT-EO  | 1250.0 | 312.5 | 0.28 (S) | MT-EO  | 1250.0 | 156.5 | 0.15 (8) |
| 1. deruginosa ATCC 27833 | Ch     | 19.5   | 0.6   |          | Sm     | 2.4    | 0.3   | 0.15 (S) |

MICa: MIC of EO or antibiotic alone; MICc ( $\mu g/mL$ ): MIC of EO or antibiotic in the most effective combination ( $\mu g/mL$ ); (S): synergy (FICI  $\leq$  0.5).

The cytotoxic efficacy of MT-EO was evaluated via the MTT assay on four human cancer cell lines: hepatocellular carcinoma (HepG2), breast cancer (MCF7) cells, lung adenocarcinoma (A-549), and colorectal carcinoma (HCT-116), alongside the non-cancerous human liver cell line LO2 [23]. Doxorubicin was employed as a positive control. The results of the 24, 48, and 72-hour exposures (Table 4 and Figure 1) showed that MT-EO had significant cytotoxic activity against all tested cell lines and exhibited dose- and time-dependent cytotoxic effects. The selectivity index of MT-EO was calculated in the range of 0.32 to 0.60 (48 h). The main compounds present in the essential oils may be responsible for the interesting cytotoxic activity, such as spathulenol [9],  $\beta$ -elemene [11],  $\delta$ -elemene [24], and bicyclogermacrene [13], which has been demonstrated to exhibit cytotoxic effects on multiple tumor cell lines. However, MT-EO exhibited weaker activity in comparison with positive control Doxorubicin.



**Figure 1**. Cytotoxic activity of MT-EO for 24 h (a); 48 h (b); 72 h (c). (P < 0.05)

 $0.33 \pm 0.01$ 

 $20.28 \pm 1.30$ 

 $0.48 \pm 0.06$ 

|       | Sample      | 24h              | 48h              | 72h              |
|-------|-------------|------------------|------------------|------------------|
|       | EO          | $40.20 \pm 2.05$ | $34.19 \pm 0.51$ | $12.41 \pm 1.24$ |
| HepG2 | Doxorubicin | $1.46 \pm 0.08$  | $1.09 \pm 0.08$  | $0.43 \pm 0.07$  |
| MCE 7 | EO          | $31.73 \pm 2.17$ | $25.20 \pm 2.28$ | $19.26 \pm 0.70$ |
| MCF-7 | Doxorubicin | $1.56 \pm 0.032$ | $0.79 \pm 0.02$  | $0.39 \pm 0.06$  |
| 1.02  | EO          | $27.67 \pm 1.95$ | $15.12 \pm 0.96$ | $12.11 \pm 1.02$ |
| LO2   | Doxorubicin | $1.80 \pm 0.29$  | $0.46 \pm 0.02$  | $0.55 \pm 0.12$  |
| A-549 | EO          | $55.56 \pm 1.96$ | $47.07 \pm 1.96$ | $40.21 \pm 2.96$ |
| A-349 | Dovorubicin | $1.04 \pm 0.07$  | $0.85 \pm 0.05$  | $0.33 \pm 0.01$  |

 $0.85 \pm 0.05$ 

 $28.89 \pm 2.19$ 

 $0.57 \pm 0.02$ 

**Table 4.** Cytotoxic activity (IC<sub>50</sub>, μg/mL) of MT-EO

Doxorubicin

Doxorubicin

The antioxidant activities of MT-EO were evaluated using three antioxidant models: DPPH, ABTS, and FRAP [25]. The results are shown in Table 5. The study confirms that MT-EO demonstrates moderate antioxidant activity in DPPH and ABTS assays, with IC<sub>50</sub> values of 434.5  $\pm$  9.6 and 98.1  $\pm$  1.1  $\mu$ g/mL, respectively. Compared to the standard antioxidants BHT and Trolox, MT-EO showed a mild free radical scavenging activity. In addition, it has a moderate activity for reducing ferric ions, with a trolox equivalent of 97.11  $\pm$  3.37 mol Trolox·g<sup>-1</sup>.

**Table 5.** Results of antioxidant activity in vitro (DPPH, ABTS and FRAP) of MT-EO

 $1.04 \pm 0.07$ 

 $41.50 \pm 2.43$ 

 $1.33 \pm 0.15$ 

| Test Sample      | DPPH IC <sub>50</sub> ( $\mu$ g/mL) <sup>a</sup> | ABTS IC50 ( $\mu g/mL$ ) <sup>a</sup> | FRAP ( $\mu$ mol Trolox × g <sup>-1</sup> ) |
|------------------|--|---------------------------------------|---|
| MT-EO            | $434.5 \pm 9.6$                                  | $98.1 \pm 1.1$                        | $97.11 \pm 3.37$                            |
| BHT <sup>b</sup> | $5.3 \pm 0.3$                                    | $2.6 \pm 0.1$                         |   |
| Trolox b         | $6.1 \pm 0.4$                                    | $5.3 \pm 0.3$                         |   |

<sup>&</sup>lt;sup>a</sup> IC<sub>50</sub> = The sample concentration for a 50% reduction in the assay; <sup>b</sup> Positive control used.

# **Supporting Information**

HCT-116

Supporting Information accompanies this paper on http://www.acgpubs.org/journal/records-ofnatural-products

# ORCID (D

Xiangyi Li: 0009-0006-2620-1486 Shu Oiu: 0009-0002-9671-5790 Shiyu Song: 0009-0009-3513-5347 Pengxiang Lai: 0000-0002-5380-4382

### References

- G.X. Xing (1999). Flora Reipublicae Popularis Sinicae. Science Press, Beijing, 4, pp.79-82. [1]
- [2] S. Mondal, H. Reddy, P. Vidya, D. Ghosh, S. Raja and S. Ganapaty (2015). Evaluations of healing potential of ethanol extract from Macrothelypteris torresiana (Gaudich) aerial parts, Int. J. Phytomed. 7, 316-323.
- J. Chen, Y. Lei, G. Wu, Y. Zhang, W. Fu, C. Xiong and J. Ruan (2012). Renoprotective potential of [3] Macrothelypteris torresiana via ameliorating oxidative stress and proinflammatory cytokines, J. Ethnopharmacol. 139, 207-213.
- W. Fang, J. Ruan, Y. Cai, A. Wei, D. Zhou and W. Zhang (2011). Flavonoids from the aerial parts of [4] Macrothelypteris torresiana, Nat. Prod. Res. 25, 36-39.
- C. Xiong, J. Ruan, Y. Tang, Y. Cai, W. Fang, Y. Zhu and D. Zhou (2009). Chromatographic fingerprint [5] analysis of Macrothelypteris torresiana and simultaneous determination of several main constituents by LC, Chromatogrography 70, 117-124.
- S. Mondal, D. Ghosh, S. Ganapaty, S.V.G. Chekubovina and M. Samal (2017). Hepatoprotective activity of [6] Macrothelypteris torresiana (Gaudich.) aerial parts against CCl<sub>4</sub>-induced hepatotoxicity in rodents and analysis of polyphenolic compounds by HPTLC, J. Pharm. Anal. 7, 181-189.

- [7] S. Mondal, D. Ghosh, S. Ganapaty, O. Manna, M.V. Reddy and V. Revanth (2016). Evaluation of analgesic, antipyretic and anti-inflammatory effects of ethanol extract from a fern species *Macrothelypteris torresiana* (Gaudich) aerial parts, *Pharmacogn. Commn.* **6**, 57-63.
- [8] X. Huang, P. Xiong, C. Xiong, Y. Cai, A. Wei, J. Wang, X. Liang and J. Ruan (2010). *In vitro* and *in vivo* antitumor activity of *Macrothelypteris torresiana* and its acute/subacute oral toxicity, *Phytomedicine* 17, 930-934.
- [9] K.F.do Nascimento, F.M.F. Moreira, J.A. Santos, C.A.L. Kassuya, J.H.R. Croda, C.A.L. Cardoso, M.D.C. Vieira, A.L.T.G. Ruiz, M.A. Foglio and J.E.de Carvalho (2018). Antioxidant, anti-inflammatory, antiproliferative and antimycobacterial activities of the essential oil of *Psidium guineense* Sw. and spathulenol, *J. Ethnopharmacol.* **210**, 351-358.
- [10] A.D.J. Dzul-Beh, K. García-Sosa, A.H. Uc-Cachón, J. Bórquez, L.A. Loyola, H.B. Barrios-García, L.M. Peña-Rodríguez and G.M. Molina-Salinas (2020). In vitro growth inhibition and bactericidal activity of spathulenol against drug-resistant clinical isolates of *Mycobacterium tuberculosis*, *Rev. Bras Farmacogn.* **29**, 798-800.
- [11] Z. Jiang, J.A. Jacob, D.S. Loganathachetti, P. Nainangu and B. Chen (2017). β-Elemene: mechanistic studies on cancer cell interaction and its chemosensitization effect, *Front. Pharmacol.* **8**, 105.
- [12] E. Sieniawska, R. Sawicki, J. Golus, M. Swatko-Ossor, G. Ginalska and K. Skalicka-Wozniak (2018). *Nigella damascena* L. essential oil-a valuable source of β-elemene for antimicrobial testing, *Molecules* 23, 256
- [13] E.B.P. da Silva, A.L. Matsuo, C.R. Figueiredo, M.H. Chaves, P. Sartorelli and J.H.G. Lago (2013). Chemical constituents and cytotoxic evaluation of essential oils from leaves of *Porcelia macrocarpa* (Annonaceae), *Nat. Prod. Commun.* **8**, 277-279.
- [14] R.P. Adams (2017). Identification of essential oil components by gas chromatography/mass spectrometry. Texensis Publishing. Gruver, Texas.
- [15] N.R. Andriamaharavo (2014). Retention Data NIST Mass Spectrometry Data Center, NIST Mass Spectrometry Data Center.
- [16] P.J. Linstrom and W.G. Mallard (2014). NIST Chemistry WebBook, NIST Standard Reference Database Number 69. (http://webbook.nist.gov).
- [17] V.I. Babushok, P.J. Linstrom and I.G. Zenkevich (2011). Retention indices for frequently reported compounds of plant essential oils, *J. Phys. Chem. Ref. Data.* **40**, 1-47.
- [18] M. Kulić, D. Drakul, D. Sokolović, J. Kordić-Bojinović, S. Milovanović and D. Blagojević (2023). Essential oil of *Satureja montana* L. from Herzegovina: assessment of composition, antispasmodic, and antidiarrheal effects, *Rec. Nat. Prod.* 17, 536-548.
- [19] Clinical and Laboratory Standards Institute (2012). Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. *Approved standard M07-A9*. National Committee for Clinical Laboratory Standards, Wayne, Pennsylvania, USA.
- [20] X.Q. Yu, W.F. He, D.Q. Liu, M.T. Feng, Y. Fang, B. Wang, L.H. Feng, Y.W. Guo and S.C. Mao (2014). A seco-laurane sesquiterpene and related laurane derivatives from the red alga *Laurencia okamurai* Yamada, *Phytochemistry* **103**, 162-170.
- [21] P. Bellio, L. Fagnani, L. Nazzicone and G. Celenza (2021). New and simplified method for drug combination studies by checkerboard assay, *MethodsX* **8**, 101543.
- [22] S. Hemaiswarya, A.K. Kruthiventi and M. Doble (2008). Synergism between natural products and antibiotics against infectious diseases, *Phytomed.* **15**, 639-652.
- [23] E. İnal, E.O. Nath, M. Abudayyak, Ş. Ulusoy, H.A. İnan, M. Çiçek and M. Kartal (2023). Chemical composition of different parts of the *Vitex agnus-castus* L. essential oils and their *in-vitro* cytotoxic activities. *Rec. Nat. Prod.* 17, 904-917.
- [24] C.Y. Xie, W. Yang, M. Li, J. Ying, S.J. Tao, K. Li, J.H. Dong and X.S. Wang (2009). Cell apoptosis induced by  $\delta$ -elemene in colorectal adenocarcinoma cells via a mitochondrial-mediated pathway, Yakugaku Zasshi **129**, 1403-1413.
- [25] B. Kiraz Kınoğlu, T. Dirmenci, S.H. Alwasel, İ. Gulcin and A. C. Goren (2023). Quantification of main secondary metabolites of *Satureja icarica* P.H. Davis (Lamiaceae) by LC-HRMS and evaluation of antioxidant capacities, *J. Chem. Metrol.* 17, 199-214.

