# **Supporting Information**

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# 3-O-Formyl-27-Hydroxyfusidic Acid: A New Metabolite of Fusidic Acid by *Cunninghamella echinulata*

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1. Experimental

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#### 1. Experimental

#### 1.1. General Procedures

Sodium fucidate was purchased from Leo Pharmaceutical Company (Ballerup, Denmark). IR and UV spectra were recorded using PerkinElmer IR and Shimadzu 60/PC ultraviolet spectrophotometers, respectively. <sup>1</sup>H and <sup>13</sup>C NMR spectra were obtained on Bruker models AMX- 500 NMR spectrometer with standard pulse sequences operating at 500 MHz for <sup>1</sup>H, and 125 MHz for <sup>13</sup>C NMR, respectively. CDCl<sub>3</sub> as solvent and tetramethyl silane (TMS) as internal standard. HRESIMS was performed with a LCT Premier XE Micromass Waters spectrometer in the positive- ionization mode (Waters Corporation). The compounds were detected on Precoated silica gel 60  $F_{254}$  plates (0.25 mm layer, E. Merck) using chloroform-methanol (5:1) or benzene-ethyl acetate-formic acid (3 mL:7 mL:1 drop) as mobile phases and visualized with *p*-anisaldehyde spray reagent after heating at 110 °C.

#### 1.2. Microorganism Strain and Culture Conditions

Biotransformation studies were performed following previously reported procedures (Ibrahim et al., 2018). The fermentation medium (pH 6.0) is composed of 1% glycerol, 1% glucose, 0.5% peptone, 0.5% yeast extract, 0.5% NaCl, and 0.5% K<sub>2</sub>HPO<sub>4</sub> in 1 L of distilled water and the medium was autoclaved at 121 °C for 15 min.

#### 1.3. Large-scale Fermentation

Biotransformation process was carried out following the published procedures [1]. Two-week old slants of *C. echinulata* were used for preparing stage I cultures of which 5 ml was inoculated into new culture media to initiate stage II cultures. After 24 h, sodium fusidate was added and incubation continued for 6 days along with organism and substrate free cultures. Cultures were filtered after acidification with 10% HCl and the metabolites were extracted from the filtrate by chloroform which was dehydrated over anhydrous sodium sulphate. TLC was carried out as discussed in general procedures.

#### 1.4. Isolation of Metabolite 2

A residue obtained by evaporating the chloroform extract (3.4 g) was loaded onto a silica gel column (300 g). Fractions of 125 ml were collected by using a gradient of ethyl acetate in benzene (0–60%) containing 0.2% formic acid which was increased to 0.4% starting from fraction 107. The residue of fractions 122-142 (300 mg), eluted with acidulated 60% ethyl acetate in benzene, was chromatographed again on a silica gel column (40 g) and 50 mL fractions were collected using a gradient of methanol in chloroform (0–10%). Fractions 37–38, eluted with 3% methanol, afforded compound **2** (10 mg).

#### 1.5. Antimicrobial Activity

Minimum inhibitory concentration of compound 2 was determined using the National Committee of Clinical Laboratory Standard and ATCC strains.

Microorganism	MIC (µg/mL)	
	1	2
Staphylococcus aureus (ATCC 25923)	0.38	1000
Escherichia coli (ATCC 25922)	-ve	2000
Pseudomonas aeruginosa (ATCC 15442)	-ve	-ve
Candida albicans (ATCC 10231)	12.5	2000

Table S1: Antimicrobial activity testing of fusidic acid (1) and the isolated metabolite 2

\* -ve at the highest tested concentration (2000  $\mu$ g/mL)



Figure S1: IR spectrum of 3-O-formyl-27-hydroxyfusidic acid (2)



Figure S2: <sup>1</sup>H NMR spectrum of 3-*O*-formyl-27-hydroxyfusidic acid (2) (CDCl<sub>3</sub>, 500 MHz)



Figure S3: <sup>13</sup>C NMR spectra of 3-*O*-formyl-27-hydroxyfusidic acid (2) (CDCl<sub>3</sub>, 125 MHz)



Figure S4: DEPT 135 spectrum of 3-O-formyl-27-hydroxyfusidic acid (2)



Figure S5: HMQC spectrum of 3-O-formyl-27-hydroxyfusidic acid (2)



**Figure S6:** HMBC spectrum of 3-*O*-formyl-27-hydroxyfusidic acid (**2**) (in CDCl<sub>3</sub> [top], and methanol-d4 [bottom])



Figure S7: COSY spectrum of 3-O-formyl-27-hydroxyfusidic acid (2)



Figure S8: NOESY spectrum of 3-O-formyl-27-hydroxyfusidic acid (2)



Figure S9: HRESIMS spectrum of 3-O-formyl-27-hydroxyfusidic acid (2) in the positive ion

mode (top). Low resolution MS in the negative ion mode (bottom).

#### References

[1] A. S. Ibrahim, K. Elokely, D. Ferreira and A. E. Ragab (2018). Microbial oxidation of the fusidic acid side chain by *Cunninghamella echinulata*, *Molecules* 23, 970-980.