

A New Diketopiperazine from the Marine Sponge *Callyspongia* Species

Bin Yang¹, Jingxia Huang², Xiuping Lin¹, Yanying Zhang¹
Huaming Tao³ and Yonghong Liu^{1*}

¹ CAS Key Laboratory of Tropical Marine Bio-resources and Ecology / Guangdong Key Laboratory of Marine Materia Medica / Research Center for Marine Microbes, South China Sea Institute of Oceanology, Chinese Academy of Sciences, Guangzhou 510301, P.R. China

² Zhongshan Ophthalmic Center, Sun Yat-Sen University, Guangzhou 510060, P.R. China

³ School of Traditional Chinese Medicine, Southern Medical University, Guangzhou 510515, P. R. China

(Received July 20, 2014; Revised September 29, 2014; Accepted October 30, 2014)

Abstract: Chemical investigation of the sponge *Callyspongia* sp. from the South China Sea afforded one new diketopiperazine, cyclo-(*R*-Pro-6-hydroxyl-*S*-Ile) (**1**), along with six known diketopiperazines: staphyloamide A (**2**), cyclo-(*S*-Pro-*S*-Phe) (**3**), cyclo-(*R*-Pro-*R*-Phe) (**4**), cyclo-(*S*-Pro-*R*-Leu) (**5**), cyclo-(*S*-Pro-*R*-Ala) (**6**), cyclo-(*R*-Tyr-*R*-Phe) (**7**), and three known tryptophan-derived alkaloids: C²- α -D-mannosylpyranosyl-tryptophan (**8**), (1*R*, 3*S*)-1-methyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylic acid (**9**), and (1*R*,3*R*)-1-methyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylic acid (**10**). The structures were determined on the basis of NMR and MS analysis, and the absolute configuration was determined by comparison of the optical rotation with the known compounds. This is the first report of compounds **1**, **2**, **8–10** from the sponge *Callyspongia*. Cyclo-(*S*-Pro-*R*-Leu) (**5**), and cyclo-(*S*-Pro-*R*-Ala) (**6**) exhibited antifouling activity against cyprid larvae of the barnacle with the LC₅₀ values of 3.5 $\mu\text{g}/\text{cm}^2$ and 6.0 $\mu\text{g}/\text{cm}^2$, respectively.

Keywords: *Callyspongia* sp.; Diketopiperazine; Chemical constituents; Antifouling activity. © 2015 ACG Publications. All rights reserved.

1. Animal Source

Callyspongia sp. belongs to the order Haplosclerida family Callyspongiidae. The genus *Callyspongia* have been found to contain a variety of structurally unique metabolites with interesting biological activities, and a number of alkaloids have been isolated from this genus so far [1-4].

The sponge was collected by hand in July 2005, off the coast of Hainan Island, P. R. China. The specimen was identified by Dr. Kyung Jin Lee, Invertebrate Research Division, National Institute of Biological Resources, Environmental Research Complex, Incheon, Korea. A voucher specimen

* Corresponding author: E- Mail: Yonghongliu@scsio.ac.cn (Y.Liu), Phone:+86-20-89023244

(0507002) was deposited with the Key Laboratory of Marine Bio-resources Sustainable Utilization, South China Sea Institute of Oceanology, Chinese Academy of Sciences.

2. Previous Studies

Diketopiperazines (DKPs) are a large and structurally varied group of natural products isolated from both terrestrial and marine organisms [5]. A number of DKPs were isolated from various marine sponges: *Calyx cf podatypa* [6], *Dysidea herbacea* [7], *Geodia baretii* [8], *Tedania ignis* [9], *Axinella* sp.[10], and *Dysidea fragilis*[11]. Many of these constituents are reported to have a broad range of bioactivities, such as antitumour, antibacterial, antifungal, antifouling, plant-growth regulatory, and other activities.[5, 12]. In our previous study, a new proline-containing dipeptide, named callyspongidi peptide A, and two known DKPs were isolated from *Callyspongia* sp.[13].

3. Present Study

The sponge (10 kg) was extracted three times with 95% EtOH (50 L) for 3 days. The extract was concentrated under reduced pressure, and partitioned between H₂O (4 L) and CHCl₃ (4 L); the CHCl₃ layer (405 g) was further partitioned between 85% EtOH (4 L) and petroleum ether (PE; 4 L) to yield 85% EtOH (98 g) and PE (270 g) fractions. The H₂O layer was partitioned between n-BuOH (4 L) and H₂O (4 L) to yield n-BuOH (71 g) fractions. Chromatographic purification of the 85% EtOH portion and n-BuOH portion yielded compounds **1–10** (Figure 1). Detailed isolation procedures for compounds **1–10** are shown in the supplementary material. Structure characterization was aided by various spectroscopic experiments such as MS, and NMR. NMR spectra were measured on Bruker AVANCE-500 spectrometer. ESI-MS was obtained from Thermo LCQ-DECA-XP LC-MS spectrometer. HR-ESI-MS data were measured on a Bruker micro TOF-QII mass spectrometer.

cyclo-(R-Pro-6-hydroxyl-R-Ile) (**1**): C₁₁H₁₈N₂O₃, White amorphous powder. $[\alpha]_D^{25}$: + 27.4 (c 0.04, MeOH). ¹H NMR (500 MHz, CD₃OD): δ 3.64-3.61 (2H, m, H-3), 3.60 (1H, d, $J = 7.5$ Hz, H-9), 2.29-2.26 (1H, m, H-5a), 2.18-2.16 (1H, m, H-4a), 2.13-2.09 (1H, m, H-5b), 2.09-2.05 (1H, m, H-10), 2.01-1.97 (1H, m, H-4b), 1.78-1.67 (1H, m, H-11a), 1.28-1.19 (1H, m, H-11b), 1.03 (3H, d, $J = 7.0$ Hz, H-13), 0.98 (3H, t, $J = 7.5$ Hz, H-12). ¹³C NMR (125 MHz, CD₃OD): δ 170.5 (C-1, C), 169.5 (C-7, C), 88.1 (C-6, C), 63.6 (C-9, CH), 46.6 (C-3, CH₂), 41.2 (C-10, CH), 38.1 (C-5, CH₂), 26.6 (C-11, CH₂), 20.3 (C-4, CH₂), 15.9 (C-13, CH₃), 11.4 (C-12, CH₃). ESI-MS (neg.): m/z 225.1 ([M – H][–]). HR-ESI-MS m/z 241.1209 (calcd for C₁₁H₁₈N₂O₃Na, 241.1210).

Antifouling Assay: Antifouling efficacies of the eleven alkaloids isolated from *Callyspongia* sp. were evaluated by the settlement inhibition assay with cyprid larvae of *Balanus reticulatus*. *B. reticulatus* adults were collected from intertidal rocks in Shenzhen, China. Antifouling efficacies of the ten alkaloids were investigated according to the method of Chen *et al.* with a minor modification [14]. The tested compounds were introduced to the glass dishes using DMSO as carrier solvent.

Antimicrobial Activity: Antimicrobial assays against three bacteria (*Bacillus subtilis*, *Staphylococcus aureus*, and *Escherichia coli*) were carried out using the disk diffusion method [15]. Chloramphenicol was used as positive controls against bacteria.

Compound **1** was obtained as white amorphous powder. Compound **1** exhibited a [M – H][–] ion peak at m/z 225.1 in the negative-ion ESI-MS. The HR-ESI-MS of **1** established its molecular formula as C₁₁H₁₈N₂O₃. Comparison of **1** with staphyloamide A (**2**) revealed that it not only exhibit the same molecular mass, but almost the same physical and NMR spectroscopic data [16], and differed mainly in the shift of the C-9 proton signals from δ_H 4.20 to 3.60. The HMBC correlations of H-9 to C-1, and C-10 and H₃-13 to C-9, C-10, and C-11 were observed. This fact described that **1** was diastereoisomer to **2** at position C-9. Such change was also observed in staphyloamide B [16] and bacillusamide B [7]. Furthermore, applying the empirical trend of the proline-containing DKPs, the optical rotation of **1** ($[\alpha]_D^{25} = + 27.4$) was allowed to propose the diketopiperazine structure cyclo-(*R*-Pro-6-hydroxyl-*S*-Ile) for compound **1** [6, 16, 17].

The identification of the other known compounds, staphyloamide A (**2**), cyclo-(*S*-Pro-*S*-Phe) (**3**) [18], cyclo-(*R*-Pro-*R*-Phe) (**4**) [6], cyclo-(*S*-Pro-*R*-Leu) (**5**) [6], cyclo-(*S*-Pro-*R*-Ala) (**6**) [19], cyclo-(*R*-Tyr-*R*-Phe) (**7**) [20], C²- α -D-mannosylpyranosyl-tryptophan (**8**) [21, 22], (1*R*, 3*S*)-1-methyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylic acid (**9**) [23, 24], and (1*R*,3*R*)-1-methyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylic acid (**10**) [23, 24], was confirmed by comparison of the ¹H and ¹³C NMR data with those reported. This is the first report of compounds **1**, **2**, **8–10** from the sponge *Callyspongia*.

The antifouling activities of compounds **1–10** against larval settlement of the barnacle *Balanus reticulatus* were evaluated. Comparison of means using ANOVA and Dunnett's test showed that compounds **5** and **6** significantly inhibited settlement compared with the negative control ($p < 0.001$) without significant toxicity ($p > 0.05$). From Dunnett's test, cyclo-(*S*-Pro-*R*-Leu) (**5**), and cyclo-(*S*-Pro-*R*-Ala) (**6**) significantly reduced larval settlement with the LC₅₀ values of 3.5 $\mu\text{g}/\text{cm}^2$ and 6.0 $\mu\text{g}/\text{cm}^2$, respectively.

MIC values for compounds **1–10** were measured against three bacteria strains of *Bacillus subtilis*, *Staphylococcus aureus*, and *Escherichia coli*. All the compounds (**1–10**) did not show antimicrobial activities (MIC > 100 $\mu\text{g}/\text{mL}$).

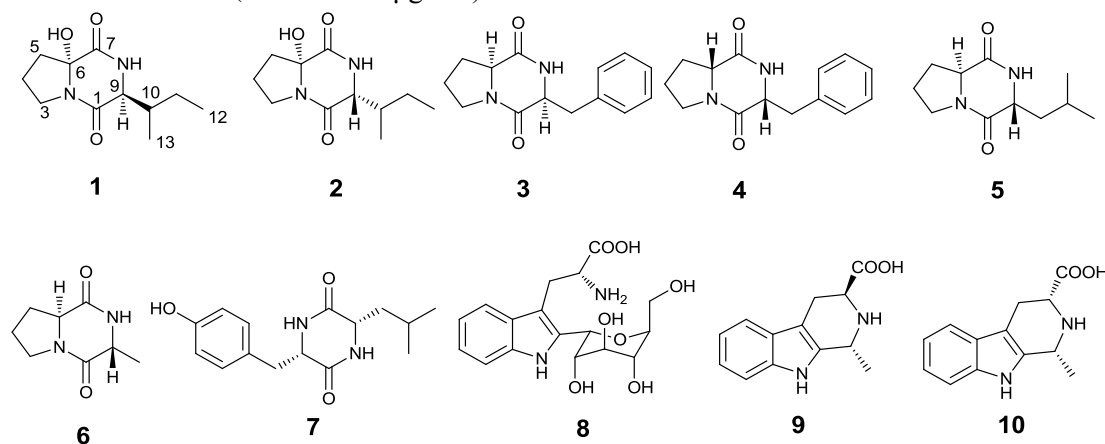


Figure 1. Structures of isolated compounds (**1–10**) from *Callyspongia* sp.

Our study revealed the chemical constituents of sponge *Callyspongia* sp., which is rich in the South China Sea. Ten compounds were isolated and purified including seven diketopiperazines and three tryptophan-derived alkaloids. Compound **1** is a new secondary metabolite. In order to detect whether these compounds play a role for ecological functions, the tests for antifouling and antibacterial activity were performed. The results showed significant toxicities against *Balanus reticulatus* larvae for compounds **5** and **6**, which suggest that they contribute to a chemical ecological function.

Acknowledgments

This study was supported by grants from the National Key Basic Research Program of China (973)'s Project (2010CB833800, 2011CB915503), the National High Technology Research and Development Program (863 Program, 2012AA092104, 2013AA092901), National Natural Science Foundation of China (21302198, 41376162, 21202080), the Chinese Academy of Sciences (XDA11030403), Guangdong Natural Science Foundation (S2012040007061).

Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/RNP>

References

- [1] P. Y. Qian, S. Dobretsov, H. U. Dahms and J. Pawlik (2006). Antifouling activity and microbial diversity of two congeneric sponges *Callyspongia* spp. from Hong Kong and the Bahamas, *Mar. Ecol. Prog. Ser.* **324**, 151–165.
- [2] R. M. Huang, W. Ma, J. D. Dong, X. F. Zhou, T. H. Xu, K. J. Lee, X. W. Yang, S. H. Xu and Y. H. Liu (2010). A new 1,4-diazepine from South China Sea marine sponge *Callyspongia* species, *Molecules* **15**, 871–877.
- [3] C. D. Pham, R. Hartmann, P. Bohler, B. Stork, S. Wesselborg, W. H. Lin, D. W. Lai and P. Proksch (2014). Callyspongiolide, a cytotoxic macrolide from the marine sponge *Callyspongia* sp., *Org. Lett.* **16**, 266–269.
- [4] F. Plisson, P. Prasad, X. Xiao, A. M. Piggott, X. C. Huang, Z. Khalil and R. J. Capon (2014). Callyspongisines A-D: bromopyrrole alkaloids from an Australian marine sponge, *Callyspongia* sp., *Org. Biomol. Chem.* **12**, 1579–1584.
- [5] R. M. Huang, X. F. Zhou, T. H. Xu, X. W. Yang and Y. H. Liu (2010). Diketopiperazines from marine organisms, *Chem. Biodivers.* **7**, 2809–2829.
- [6] M. Adamczeski, A. R. Reed and P. Crews (1995). New and known diketopiperazines from the Caribbean sponge, *Calyx cf podatypa*, *J. Nat. Prod.* **58**, 201–208.
- [7] R. Kazlauskas, P. T. Murphy and R. J. Wells (1978). Diketopiperazine derived from trichloroleucine from sponge *Dysidea-herbacea*, *Tetrahedron Lett.* **49**, 4945–4948.
- [8] J. Bergman (2013). Synthesis and studies of two marine indole alkaloids, baretin and caulersin, *Phytochem. Rev.* **12**, 487–494.
- [9] F. J. Schmitz, D. J. Vanderah, K. H. Hollenbeak, C. E. L. Enwall, Y. Gopichand, P. K. Sengupta, M. B. Hossain and D. Vanderhelm (1983). Metabolites from the marine sponge *Tedania-ignis*-A new atisanediol and several known diketopiperazines, *J. Org. Chem.* **48**, 3941–3945.
- [10] R. M. Huang, T. Yan, Y. Peng, X. F. Zhou, X. W. Yang and Y. H. Liu (2014). Diketopiperazines from the marine sponge *Axinella* sp., *Chem. Nat. Compd.* **50**, 191–193.
- [11] J. Y. Su, Y. L. Zhong, L. M. Zeng, S. Wei, Q. W. Wang, T. C. W. Mak and Z. Y. Zhou (1993). 3 new diketopiperazines from a marine sponge *Dysidea-fragilis*, *J. Nat. Prod.* **56**, 637–642.
- [12] M. B. Martins and I. Carvalho (2007). Diketopiperazines: biological activity and synthesis, *Tetrahedron* **63**, 9923–9932.
- [13] B. Yang, J. D. Dong, X. F. Zhou, X. W. Yang, K. J. Lee, L. S. Wang, S. Zhang and Y. H. Liu (2009). Proline-containing dipeptides from a marine sponge of a *Callyspongia* species, *Helv. Chim. Acta.* **92**, 1112–1117.
- [14] J. D. Chen, R. Z. Yi, Y. M. Lin, D. Q. Feng, H. C. Zhou and Z. C. Wang (2011). Characterization of terpenoids from the root of *Ceriops tagal* with antifouling activity, *Int. J. Mol. Sci.* **12**, 6517–6528.
- [15] S. K. S. Al-Burtamani, M. O. Fatope, R. G. Marwah, A. K. Onifade and S. H. Al-Saidi (2005). Chemical composition, antibacterial and antifungal activities of the essential oil of *Haplophyllum tuberculatum* from Oman, *J. Ethnopharmacol.* **96**, 107–112.
- [16] A. I. M. Khedr, I. Kouno, T. Tanaka and K. Yamada (2013). New diketopiperazine derivatives from culture broth of *Staphylococcus* sp. isolated from *Corallina Officinalis lineaus*, *Heterocycles* **87**, 1029–1037.
- [17] K. Yonezawa, K. Yamada and I. Kouno (2011). New diketopiperazine derivatives isolated from sea urchin-derived *Bacillus* sp., *Chem. Pharm. Bull.* **59**, 106–108.
- [18] F. Fdhila, V. Vazquez, J. L. Sanchez and R. Riguera (2003). DD-diketopiperazines: Antibiotics active against *Vibrio anguillarum* isolated from marine bacteria associated with cultures of *Pecten maximus*, *J. Nat. Prod.* **66**, 1299–1301.
- [19] K. Suguna, S. Ramakumar and K. D. Kopple (1984). Structure of cyclo(-L-Leucyl-L-Tyrosyl-) monohydrate, C₁₅H₂₀N₂O₃·H₂O, *Acta. Crystallogr. C.* **40**, 2053–2056.
- [20] Y. Takaya, T. Furukawa, S. Miura, T. Akutagawa, Y. Hotta, N. Ishikawa and M. Niwa (2007). Antioxidant constituents in distillation residue of *Awamori spirits*, *J. Agr. Food Chem.* **55**, 75–79.
- [21] R. J. Capon and N. S. Trotter (2005). N-3,5 '-cyclooxanthosine, the first natural occurrence of a cyclonucleoside, *J. Nat. Prod.* **68**, 1689–1691.

- [22] A. Garcia, L. A. Lenis, C. Jimenez, C. Debitus, E. Quinoa and R. Riguera (2000). The occurrence of the human glycoconjugate C-2-alpha-D-mannosylpyranosyl-L-tryptophan in marine ascidians, *Org. Lett.* **2**, 2765–2767.
- [23] T. Herraiz (1997). Analysis of tetrahydro-beta-carbolines and their precursors by electron ionization mass spectrometry. Identification in foodstuffs by gas chromatography mass spectrometry, *Rapid. Commun. Mass. Sp.* **11**, 762–768.
- [24] F. M. Kuo, M. C. Tseng, Y. H. Yen and Y. H. Chu (2004). Microwave accelerated Pictet-Spengler reactions of tryptophan with ketones directed toward the preparation of 1,1-disubstituted indole alkaloids, *Tetrahedron* **60**, 12075–12084.

ACG
publications

© 2015 ACG Publications