

Chemical Constituents, Antimicrobial and Cytotoxic Activities of *Hypericum riparium* (Guttiferae)

Michel F. Tala^{1,2}, Patricia D. Tchakam³, Hippolyte K. Wabo^{1*}, Ferdinand
M. Talontsi², Pierre Tane¹, Jules R. Kuate³, Léon A. Tapondjou¹ and
Hartmut Laatsch²

¹ Department of Chemistry, University of Dschang, P.O. Box 67, Dschang, Cameroon

² Institute of Organic and Biomolecular Chemistry, University of Göttingen, Tammannstrasse 2, D-
37077 Göttingen, Germany

³ Department of Biochemistry, University of Dschang, P.O. Box 67, Dschang, Cameroon

(Received May 24, 2012; Revised August 2, 2012; Accepted September 27, 2012)

Abstract: Betulinic acid (**1**), 5-hydroxy-3-methoxyxanthone (**2**), 1,6-dihydroxy-7-methoxyxanthone (**3**), daucosterol (**4**), bijaponicaxanthone C (**5**), hypercalin C (**6**), 1-hydroxy-6,7-dimethoxyxanthone (**7**), cadensin D (**8**) and 5-hydroxy-1,3-dimethoxyxanthone (**9**) were isolated from the roots of *Hypericum riparium*. These compounds are reported for the first time from this plant. The extracts and two of the isolated compounds (**2** and **8**) exhibited both antibacterial and antifungal activities that varied between the microbial species (MIC = 0.97-250 µg/mL). In addition, the brine-shrimp (*Artemia salina*) lethality bioassay of compound **6** showed potent cytotoxicity with LD₅₀ of 3.23 µg/mL.

Keywords: *Hypericum riparium*; antibacterial; antifungal; cytotoxic.

1. Plant source

The genus *Hypericum* occurs widely in temperate regions and have been used since ancient times as folk remedies and credited with a long list of medicinal uses, including antiviral, antimicrobial, antifungal, antitumor, analgesic, anxiolytic, sedative and for the treatment of neurological disorders and depression [1]. These effects have inspired investigations of secondary metabolites from *Hypericum* species. As results, various classes of bioactive components mainly xanthenes, phloroglucinols, flavonoids, naphthodianthrones and essential oils have been isolated and identified [1-5]. In continuation of our investigations on plants of *Hypericum* genus [6,7] we herein report the isolation of nine compounds from the roots of *Hypericum riparium*, together with the antimicrobial and cytotoxic activities of extracts and some isolated compounds. The roots of *H. riparium* A. Chev. were collected in June 2010 at Mount Bamboutos, West Region of Cameroon. Identification was done at the Cameroon National Herbarium, Yaounde, where a voucher specimen (No 33796 HNC) has been deposited.

* Corresponding author : E-Mail : [hksamdemw@yahoo.ca](mailto:hkamdemw@yahoo.ca); Phone: +237 77 74 07 53; Fax: +237 33 45 17 35

2. Previous studies

To the best of our knowledge, there are no phytochemical and biological reports on *H. riparium*.

3. Present study

Chromatography of the EtOAc-soluble portion of the CH₂Cl₂-MeOH (1:1) extract of the roots of *H. riparium* afforded betulinic acid (**1**), 5-hydroxy-3-methoxyxanthone (**2**), 1,6-dihydroxy-7-methoxyxanthone (**3**), hypercalin C (**6**) and cadensin D (**8**). EtOAc-insoluble residue was subjected to repeated column chromatography to yield daucosterol (**4**), bijaponicaxanthone C (**5**), 1-hydroxy-6,7-dimethoxyxanthone (**7**) and 5-hydroxy-1,3-dimethoxyxanthone (**9**).

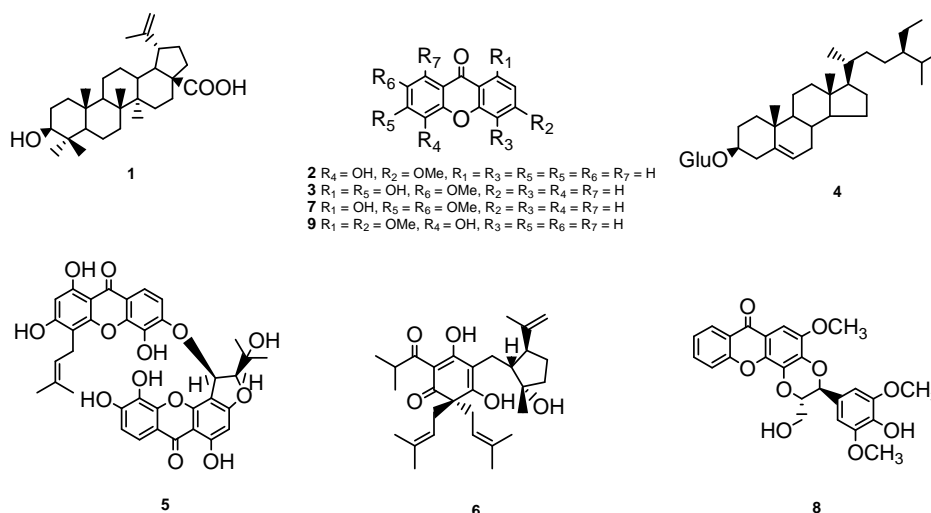


Figure 1. Compounds **1-9** isolated from *H. riparium*

Cytotoxicity assay [8]: The EtOAc extract and compound **6** showed potent cytotoxic activity in the brine-shrimp (*Artemia salina*) lethality bioassay. It was evident from the results that compound **6** was significantly lethal with LD₅₀ value of 3.23 µg/mL (Table 1). The other isolated compounds did not show any activity in this test.

Table 1. Cytotoxic activity of the EtOAc extract and compound **6** against *Artemia salina*^a

	Doses (µg/mL)	3.12	6.25	10.0	12.5	25	50	100	LD ₅₀ (µg/mL)
% Deaths	EtOAc extract	0	0	100	100	100	100	100	<10
	6	0	47	100	100	100	100	100	3.23
	Actinomycin D ^b	100	100	100	100	100	100	100	0.02

^a 10 µL of 10 µg/mL for each compound was tested. Mortality in % is determined after 24 h. 100% mortality: high active sample; ^b Reference drug

Antimicrobial activities [9]: The crude CH₂Cl₂-MeOH (1:1) extract, EtOAc-soluble fraction, EtOAc-insoluble fraction as well as compounds **2** and **8** exhibited both antibacterial and antifungal activities that varied between the microbial species (MIC = 0.97-250 µg/mL) (Table 2). The EtOAc extract was more active than the crude extract and the EtOAc-insoluble portion. Compound **2** has a large spectrum of activity whereas compound **8** showed a strong activity against *Candida lusitanae* (MIC = 3.90 µg/mL), compared to that of the reference drug (MIC = 7.81 µg/mL). Compound **6** was inactive against all the tested microorganisms. This is the first report on the antibacterial and antifungal activities of compound **2** and the antifungal activity of compound **8** against *Candida lusitanae*. Compound **2** was recently found to show a weak activity on the multidrug-resistant W2mef laboratory strain, and a field isolate (SHF4) of *Plasmodium falciparum* [7].

Table 2. Inhibition parameters (MIC, MBC and MFC) of the extracts and compounds **2**, **6** and **8** ($\mu\text{g/mL}$)

Microorganism	Parameters	Tests substances						Reference antibiotics ^c
		CE	EASF	EAI F	2	6	8	
Bacteria								
<i>Staphylococcus aureus</i>	MIC	1000	31.5	500	-	-	-	15.32
	MBC	-	250	-	-	-	-	15.32
<i>Escherichia coli</i>	MIC	500	125	125	-	-	125	1.95
	MBC	-	1000	500	-	-	-	3.9
<i>Shigella flexneri</i>	MIC	1000	1000	-	0.97	-	-	1.95
	MBC	-	-	-	1.95	-	-	7.81
<i>Salmonella typhi</i>	MIC	1000	15.62	62.5	1.95	-	-	1.95
	MBC	-	250	250	1.95	-	-	1.95
<i>Proteus mirabilis</i>	MIC	250	62.5	62.5	62.5	-	-	3.90
	MBC	-	250	250	62.5	-	-	7.81
<i>Enterococcus faecalis</i>	MIC	-	1000	-	0.97	-	125	1.95
	MBC	-	-	-	1.95	-	-	1.95
Yeast								
<i>Candida lusitaniae</i>	MIC	250	500	1000	-	-	3.90	7.81
	MFC	250	1000	-	-	-	3.90	15.32
<i>Cryptococcus neoformans</i>	MIC	250	-	250	250	-	-	1.95
	MFC	-	-	-	250	-	-	3.90
<i>Candida glabrata</i>	MIC	250	-	-	-	-	-	1.95
	MFC	1000	-	-	-	-	-	3.90
<i>Candida parapsilosis</i>	MIC	125	1000	250	-	-	-	31.25
	MFC	1000	-	-	-	-	-	-
<i>Candida krusei</i>	MIC	-	-	-	-	-	-	31.25
	MFC	-	-	-	-	-	-	-

-. Not active ($>1000 \mu\text{g/mL}$ for crude extracts, $>250 \mu\text{g/mL}$ for compounds tested on yeasts and $125 \mu\text{g/mL}$ for compounds tested on bacteria); c: Ciprofloxacin for bacteria and nystatin for yeasts; CE: crude CH_2Cl_2 -MeOH (1:1) extract; EASF: EtOAc-soluble fraction; EAI: EtOAc-insoluble fraction

Chemotaxonomic significance: The present study reports the isolation of four xanthenes (**2**, **3**, **7** and **9**), one prenylated bisxanthone (**5**), and one xanthonolignoid (**8**) together with one phloroglucinol (**6**), one triterpene (**1**) and one steroid (**4**) for the first time from the roots of *H. riparium*. Oxygenated xanthenes, xanthonolignoids and phloroglucinol are common to the Guttiferae family and particularly in the genus *Hypericum* [3-6]. 1-Hydroxy-6,7-dimethoxyxanthone (**7**) and cadensin D (**8**) have been reported from *H. perforatum* and *H. subalatum* respectively [4,5]. This is the second report of compounds **5** and **6** in the genus. Bijaponicaxanthone C (**5**) has been previously isolated from *H. japonicum* [10], while hypercalin C (**6**) was obtained from *H. calycinum* [11]. Interestingly, 1,6-dihydroxy-7-methoxyxanthone (**3**) and 5-hydroxy-1,3-dimethoxyxanthone (**9**) are reported here for the first time from the genus *Hypericum*. Compound **9** has been isolated from the genera *Kielmeyera* and *Garcinia* of the same family [12,13], while compound **3** was isolated only once from *Poeciloneuron pauciflorum* [14]. We have recently reported the isolation and characterization of several xanthenes including 5-hydroxy-3-methoxyxanthone (**2**) from the stem bark of *H. lanceolatum* [6]. *H. riparium* is one of the six *Hypericum* species found in Cameroon [6]. Thus, isolation of compounds **2**, **3** and **9** in the present investigation might be a useful contribution to the chemotaxonomic studies of the Cameroonian *Hypericum* species as well as of the Guttiferae family.

Acknowledgments

This research was supported by the International Foundation for Science, Stockholm, Sweden, and the Organization for the Prohibition of Chemical Weapons, The Hague, Netherlands (IFS-OPCW, Grant No F/4901-1) and the German Academic Exchange Service (DAAD) (Germany) through a travel grant to M.F.T. at the Institute of Organic and Biomolecular Chemistry at the University of Göttingen.

Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/RNP>

References

- [1] A. Pinarosa (2005). A survey on the *Hypericum* genus: secondary metabolites and bioactivity, *Stud. Nat. Prod. Chem.* **30**, 603-634.
- [2] B. Williams, J. Hoch, T. E. Glass, R. Evans, J. S. Miller, J. H. Wisse and D. G. I. Kingston (2003). A novel cytotoxic guttiferone analogue from *Garcinia macrophylla* from the Suriname Rainforest, *Planta Med.* **64**, 864-866.
- [3] N. Tanaka and Y. Takaishi (2006). Xanthenes from *Hypericum chinense*, *Phytochemistry.* **67**, 2146-2151.
- [4] F. Ferrari, G. Pasqua, B. Monacelli, P. Cimino and B. Botta (2005). Xanthenes from calli of *Hypericum perforatum* subsp. *perforatum*, *Nat. Prod. Res.* **19**, 171-176.
- [5] M. T. Chen (1988). Xanthonolignoids from *Hypericum sublatum*, *Heterocycles* **27**, 2589-2594.
- [6] H. K. Wabo, T. K. Kowa, A. H. N. Lonfouo, A. T. Tchinda, P. Tane, H. Kikuchi, M. Frédérick and Y. Oshima (2012). Phenolic compounds and terpenoids from *Hypericum lanceolatum*, *Rec. Nat. Prod.* **6**, 94-100.
- [7] D. Zofou, T. K. Kowa, H. K. Wabo, M. N. Ngemenya, P. Tane and V. P. K. Titanji (2011). *Hypericum lanceolatum* (Hypericaceae) as a potential source of new anti-malarial agents: a bioassay-guided fractionation of the stem bark, *Malar. J.* **10**, 167, 1- 7.
- [8] R. M. Ngoumfo, J.-B. Jouda, F. T. Mouafo, J. Komguem, C. D. Mbazona, T. C. Shiao, M. I. Choudhary, H. Laatsch, J. Legault, A. Pichette and R. Roy (2010). In vitro cytotoxic activity of isolated acridones alkaloids from *Zanthoxylum leprieurii* Guill. et Perr, *Bioorg. Med. Chem.* **18**, 3601-3605.
- [9] J. D. Tamokou, M. F. Tala, H. K. Wabo, J. R. Kuate and P. Tane (2009). Antimicrobial activities of methanol extract and compounds from stem bark of *Vismia rubescens*, *J. Ethnopharmacol.* **124**, 571-575.
- [10] P. Fu, W. D. Zhang, T. Z. Li, R. H. Liu, H. L. Li, W. Zhang and H. S. Chen (2005). A new bisxanthone from *Hypericum japonicum* Thunb. Ex Murray, *Chin. Chem. Lett.* **16**, 771-773.
- [11] L. A. Decosterd, H. Stoeckli-Evans, J.-C. Chapuis, B. Sordat and K. Hostettmann. (1989). New cell growth-inhibitory cyclohexadienone derivatives from *Hypericum calycinum* L., *Helv. Chim. Acta.* **72**, 1833-1845.
- [12] D. B. Orrea, G. L. F. e Silva, O. R. Gottlieb and S. J. Goncalves (1970). Chemistry of Brazilian Guttiferae. XVIII. Quinone and xanthone constituents of *Kielmeyera rupestris*, *Phytochemistry.* **9**, 447-51.
- [13] F.-F. Hong, Y. Chen and G.-Z. Yang (2008). Chemical constituents from the bark of *Garcinia xanthochymus* and their 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical-scavenging activities, *Helv. Chim. Acta.* **91**, 1695-1703.
- [14] H. Tosa, M. Iiunuma, K.-I. Murakami, T. Ito, T. Tanaka, V. Chelladurai and S. Riswan (1997). Three xanthenes from *Poeciloneuron pauciflorum* and *Mammea acuminata*, *Phytochemistry.* **45**, 133-136.

A C G
publications

© 2013 Reproduction is free for scientific studies