

Litsine A: A New Aporphine Alkaloid from the Root Barks of *Litsea glutinosa*

Yan Jin^{1,2}, Younan Wu^{1,2}, Yiyi Li³, Caiyun Zhang², Wanying Sun^{1,2},
Lin Dong^{*1,2} and Xiaopo Zhang^{*1,2}

¹ Hainan Provincial Key Laboratory of R&D on Tropical Herbs, Hainan Medicinal University,
Haikou 571199, P.R.China

² School of Pharmaceutical Sciences, Hainan Medicinal University, Haikou 571199, P.R.China

³ School of Basic Medicine and Life Science, Hainan Medicinal University, Haikou 571199,
P.R.China

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Abstract: A new aporphine alkaloid, named Litsine A, along with two known ones were isolated from the root barks of *Litsea glutinosa*. The new structure was determined by various spectroscopic techniques including 1D(¹H-, ¹³C-NMR), 2D-NMR(HMBC, HSQC, COSY and ROESY) and high resolution electrospray ionization mass spectrometry (HR-ESI-MS), where as the structures of the known compounds were elucidated through 1D-NMR, mass spectrometric analysis and compared with the reported data in literature. The effects of these compounds on glucose uptake in C2C12 myotubes were evaluated at the concentration of 10μM. The three compounds exhibited potent activity, with **1** as the best one, in increasing glucose uptake.

Keywords: *Litsea glutinosa*; Litsine A; aporphine alkaloids; glucose uptake activities. © 2018 ACG Publications. All rights reserved.

1. Plant Source

In the ongoing search of phytochemical studies of anti-diabetic plants distributed in Hainan Island, China. Herein, we report the isolation, structure elucidation and biological studies of a new aporphine alkaloid, Litsine A (**1**) along with two known compounds, boldine (**2**) [1] and laurilitsine (**3**)[2] (Figure 1), from the root barks of *L. glutinosa*. The investigated plant was collected from Wenchang City of Hainan Province, China, in October 2017, and authenticated by Prof. Niankai Zeng (School of Pharmaceutical Science, Hainan Medical University) according to the Flora of China. A voucher specimen (No.LG201710) was deposited at the herbarium of School of Pharmaceutical Science, Hainan Medical University.

* Corresponding authors: E- Mail: 729907108@qq.com (Lin Dong), Phone +86-898-5337; z_xp1412@163.com (Xiaopo Zhang), Phone +86+898-3826.

2. Previous Studies

L. glutinosa is a member of the genus *Litsea* in the Lauraceae family, which is widely distributed in subtropical and tropical regions. In China, this plant has been used as traditional medicine for treatment of many diseases [3,4]. Pharmacological investigations indicated that the leaves and barks of *L. glutinosa* possessed anti-bacterial, anti-inflammatory, anti-nociceptive, analgesic, and anti-diabetic properties [5,6]. Studies on chemical constituents have led to the isolation of lignans, alkaloids, etc [7,8]. Previously, we reported three new lignan glycosides and other known compounds from this plant [9,10].

3. Present Study

The root barks of *L. glutinosa* (7.0 kg) were air-dried, cut into pieces, and were extracted with ethanol. The ethanol extract was concentrated under reduced pressure to give a residue (660 g). The residue was dissolved in water, and then its pH value was adjusted to 2 by adding 1% H₂SO₄. The acidic solution was partitioned by chloroform to remove the lipid-soluble compounds. Then the pH value of the residue was adjusted to 10 by adding ammonium hydroxide. The pH of the ammonium hydroxide is 12, and the concentration is 25%. Then, ethyl acetate was used to extract the residue to give the alkaloid-rich extract (55 g) after concentration. The alkaloid-rich extract was separated by silica gel column chromatography using a gradient ratio of dichloromethane-acetone as the eluent to give five fractions (Fra1-Fra5). Fra.3 was further purified by Sephadex LH-20 to afford four fractions (subfra1-subfra4). Subfra.2 was purified by HPLC using a mixture of methanol-water (45:55) to afford Litsine A (**1**, 15 mg). Subfra3 was isolated by HPLC using a mixture of methanol-water (45:55) to give boldine (**2**, 10 mg) and laurilitine (**3**, 8 mg).

Litsine A (1): Yellow powder (MeOH), $[\alpha]_{25}^D = 35$ (c 0.05), UV (MeOH) λ_{max} (log ϵ) 258 nm. ¹H NMR (600 MHz, CD₃OD) δ (ppm) = 9.50 (1H, s, CHO), 7.95 (1H, s, H-11), 7.37 (1H, d, $J = 6.0$ Hz, H-2'), 6.78 (1H, s, H-8), 6.65 (1H, d, $J = 6.0$ Hz, H-3'), 6.52 (1H, s, H-3), 4.18 (1H, d, $J = 12.0$ Hz, H-5'a), 4.01 (1H, d, $J = 12.0$ Hz, H-5'b), 3.86 (3H, s, 10-OMe), 3.57 (3H, s, 1-OMe), 3.18 (1H, m, H-5 α), 3.16 (1H, m, H-7 α), 3.15 (1H, m, H-6 α), 2.98 (1H, m, H-4 α), 2.62 (1H, m, H-4 β), 2.60 (1H, m, H-5 β), 2.49 (1H, m, H-7 β). ¹³C NMR (150 MHz, CD₃OD) δ (ppm) = 144.9 (C, C-1), 151.1 (C, C-2), 115.6 (CH, C-3), 128.7 (C, C-3a), 131.3 (C, C-3b), 30.3 (CH₂, C-4), 51.9 (CH₂, C-5), 60.8 (CH, C-6a), 35.4 (CH₂, C-7), 127.5 (C, C-7a), 116.4 (CH, C-8), 147.7 (C, C-9), 148.4 (C, C-10), 113.3 (CH, C-11), 131.4 (C, C-11a), 130.8 (C, C-11b), 154.6 (C, C-1'), 125.2 (CH, C-2'), 114.2 (CH, C-3'), 160.4 (C, C-4'), 52.0 (CH₂, C-5'), 61.1 (CH₃, 1-OMe), 57.1 (CH₃, 10-OMe), 179.8 (CH, 1'-CHO). HR-ESI-MS: m/z 422.1599 ($[M + H]^+$, calcd. C₂₄H₂₄NO₆ for 422.1604). ESI(+)-MS: m/z 422.1 $[M+H]^+$, 297.1, 284.1, 265.1, 109.2 (C₆H₅O₂⁺).

Cell culture: C2C12 myoblasts were maintained in DMEM supplemented with 10% FBS at 37 °C and 5% CO₂. To induce differentiation, media was replaced with DMEM containing 2% horse serum when the cells reached confluence. The C2C12 cells were kept in this differentiating medium for 5 days, which allowed cells to be fully differentiated and used in the following experiments.

Glucose uptake assay: Glucose uptake assay was performed as previously reported [11]. Briefly, differentiated C2C12 myotubes were incubated with the serum-free DMEM containing the fluorescent glucose analog 2-NBDG (10 μ M) and various compounds (10 μ M) or insulin (100 nM). After incubation for 12 h, medium was removed and cells were washed with phosphate-buffered saline (PBS) twice. Cells were scraped out in 1 mL of PBS and transferred into 5 mL polystyrene round-bottom tubes (BD Falcon) and kept at 4 °C. The amount of 2-NBDG taken up by the cells was measured by determining the fluorescence intensities at Ex/Em = 475 nm/550 nm using a Tecan Infinite M1000Pro Microplate Reader (TECAN Group Ltd, Shanghai, China).

The root barks of *L. glutinosa* were air-dried, cut into pieces, and were extracted with ethanol-water. The ethanol extract was concentrated under reduced pressure to give a residue. The residue was dissolved in water, and then its pH value was adjusted to 2 by adding 1% H₂SO₄. The acidic solution was partitioned by chloroform to remove the lipid-soluble compounds. Then the pH value of the residue was adjusted to 10 by adding ammonia. Then, ethyl acetate was used to extract the basic solution, after concentration, to give the alkaloid-rich extract. Phytochemical investigation on alkaloid-rich extract has resulted in the isolation of one new aporphine alkaloid, Litsine A (**1**) (Figure 1).

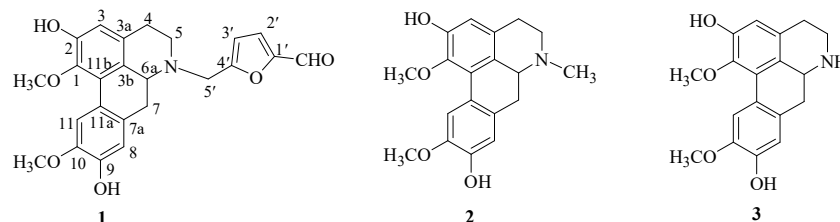


Figure 1. Structure of compounds **1-3** isolated from *L. glutinosa*

Compound **1**, Litsine A, was obtained as yellow powder. Its molecular formula was established to be C₂₄H₂₃NO₆ by interpretation of HRESIMS, requiring 14 degrees of unsaturation. The ¹H NMR spectrum showed notable signals attributing to two methoxy protons at δ_{H} 3.57 (3H, s), 3.86 (3H, s). Further examination of the spectrum revealed the existence of one aldehyde proton at $\delta = 9.50$ (1H, s) and a set of aromatic protons at $\delta = 6.65$ (1H, d, $J = 6.0$ Hz, H-3'), 7.37 (1H, d, $J = 6.0$ Hz, H-2'), 6.52 (1H, s, H-3), 6.78 (1H, s, H-8), 7.95 (1H, s, H-11). Moreover, the two deshielded methylene protons at $\delta = 3.18$ (1H, m, H-5 α), 2.60 (1H, m, H-5 β), 4.18 (1H, d, $J = 12.0$ Hz, H-5'a), 4.01 (1H, d, $J = 12.0$ Hz, H-5'b), and a methine proton at δ_{H} 3.16 (1H, m) were consistent with the presence of *N*-methylene and *N*-methine. Its ¹³C NMR alongside HSQC data showed signals for two methoxy groups, four methylenes, six methines, one aldehyde carbon, and eleven non-protonated carbons.

The ¹H-¹H COSY and HSQC spectra of **1** revealed the existence of three units as shown in Figure 2. The HMBC spectrum of **1** exhibited correlations from the aromatic proton at H-3 (δ_{H} 6.52) to carbons at δ_{C} 151.1 (C-2), δ_{C} 144.9 (C-1), δ_{C} 128.7 (C-3a) and δ_{C} 30.3 (C-4) and from the methylene proton at δ_{H} 3.18 (H-5) to carbons at δ_{C} 30.3 (C-4), δ_{C} 128.7 (C-3a) and δ_{C} 60.8 (C-6a). The correlations from methylene protons at δ_{H} 3.16, 2.49 (H-7) to carbons at δ_{C} 60.8 (C-6a), δ_{C} 131.3 (C-3b), δ_{C} 127.5 (C-7a) and δ_{C} 131.4 (C-11a). The correlations from H-8 (δ_{H} 6.78) to C-7 (δ_{C} 35.4), C-7a (δ_{C} 127.5), C-9 (δ_{C} 147.7) and C-10 (δ_{C} 148.4) and the correlations from H-11 (δ_{H} 7.95) to C-10 (δ_{C} 148.4), C-9 (δ_{C} 147.7), C-11a (δ_{C} 131.4) and C-11b (δ_{C} 130.8) verified the connection from C-7 to C-11. These data indicated that **1** had an aporphine alkaloid skeleton similar to that of boldine [1,10]. However, some differences in the NMR signals for the side chain were observed. The HMBC spectrum of **1** exhibited correlations from H-3' (δ_{H} 6.65) to C-1' (δ_{C} 154.6), C-2' (δ_{C} 125.2) and C-4' (δ_{C} 160.4) suggested the presence of a furan moiety and the aldehyde group was attached to C-1' (δ_{C} 154.6). Its ESI-MS spectrum suggested the presence of the furanoaldehyde moiety (m/z 109.2). The side-chain connectivity was deduced from the HMBC correlations from H-5' (δ_{H} 4.01, 4.18) to C-5 (δ_{C} 51.9), C-6a (δ_{C} 60.8), C-4' (δ_{C} 160.4) and C-3' (δ_{C} 114.2). These HMBC correlations also displayed that the furan moiety was attached to C-5' and C-5' was connected to the *N* atom. Finally, the correlations from methoxy proton δ_{H} 3.57 (3H, s) to C-1 (δ_{C} 144.9) and correlations from the methoxyl proton δ_{H} 3.86 (3H, s) to C-10 (δ_{C} 148.4) together with the ROESY correlations between the protons at δ_{H} 7.95 (H-11) and δ_{H} 3.57 (1-OCH₃) and δ_{H} 3.86 (10-OCH₃) suggested the two methoxy groups attached to C-1 and C-10, respectively. Thus, compound **1** was assigned as a new aporphine alkaloid with a given name Litsine A, as illustrated in Figure 1.

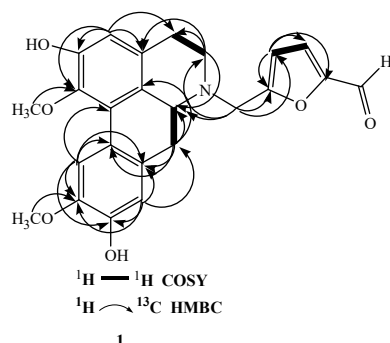


Figure 2. Key ^1H - ^1H COSY and HMBC correlations of compound **1**

Compounds **1-3** were tested for their effects on stimulating glucose uptake in C2C12 myotubes. These results showed that the three compounds increased the glucose uptake potently and **1** exhibited the best activity as shown in Figure 3.

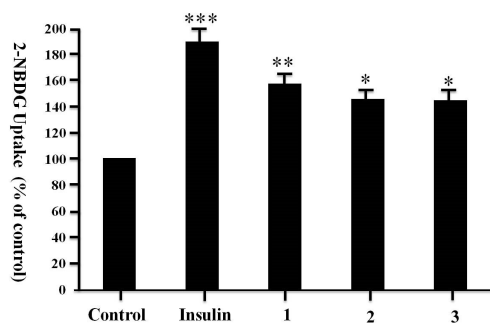


Figure 3. Effects of compounds **1-3** on glucose uptake in C2C12 myotubes

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Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/journal/records-of-natural-products>

ORCID

Yan Jin: [0000-0002-3298-1822](https://orcid.org/0000-0002-3298-1822)

Younan Wu: [0000-0003-4524-815X](https://orcid.org/0000-0003-4524-815X)

Yiying Li: [0000-0001-6855-522X](https://orcid.org/0000-0001-6855-522X)

Caiyun Zhang: [0000-0002-4432-5320](https://orcid.org/0000-0002-4432-5320)

Wanying Sun: [0000-0002-1157-5489](https://orcid.org/0000-0002-1157-5489)

Lin Dong: [0000-0003-0877-349X](https://orcid.org/0000-0003-0877-349X)

Xiaopo Zhang: [0000-0003-2614-7425](https://orcid.org/0000-0003-2614-7425)

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