

Tigliane-Type Diterpenoids from the Seeds of *Croton tiglium*

Lijuan Zhang ¹, Fei Li ², Jianyong Zhu ³ and Qian Niu ^{1*}

¹Department of Pharmacy, Bozhou Vocational and Technical College, Bozhou 236800, P.R. China

²Bozhou City Food and Drug Inspection Center, Bozhou 236800, P.R. China

³Clinical Laboratory Medicine Center, Yueyang Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai 200437, P.R. China

(Received May 07, 2022; Revised July 17, 2022; Accepted July 19, 2022)

Abstract: A new tigliane-type diterpenoid (**1**) and three known analogues (**2–4**) were isolated from the seeds of *Croton tiglium*. Extensive spectroscopic analyses, especially the 2D NMR experiments were used to determine their structures. The absolute configuration of **1** was defined by single-crystal X-ray diffraction data. The cytotoxicity **1–4** was evaluated against melanoma cell line A375, and the results showed that compounds **1**, **3**, and **4** exhibited certain cytotoxicities.

Keywords: *Croton tiglium*; tigliane-type diterpenoid; cytotoxicity. © 2022 ACG Publications. All rights reserved.

1. Introduction

Tigliane diterpenoids are a class of diterpenoids with a 5/7/6/3-fused carbon skeleton. Most of this type compounds were mainly found in Euphorbiaceae and Thymelaeaceae plants, and were reported to possess a wide range of biological properties [1]. *Croton tiglium* L. belongs to Euphorbiaceae family, and the seeds of this plant are well known as “Ba Dou” in traditional Chinese medicine to treat evil sores and scabies [2]. Previous studies have shown that tigliane diterpenoids were the main components in this plant, and many of these compounds exhibit diverse bioactivities, including cytotoxic, antiviral and anti-inflammatory activities [3–8]. In the present study, a new tigliane diterpenoid (**1**) and three known analogues (**2–4**) were obtained from the seeds of *C. tiglium* (Figure 1). The structures were characterized by spectroscopic data analyses. Herein, the isolation and structural identification of compounds **1–4**, as well as their cytotoxicities are reported.

2. Materials and Methods

2.1. General Experimental Procedures

X-ray data were obtained via a Bruker APEX-II CCD diffractometer, and the melting point was measured by an X-4 melting instrument. UV spectrum were obtained using Shimadzu UV-2450

* Corresponding author: E-Mail: 0430070164@bzy.edu.cn

spectrophotometry. Optical rotation data were obtained by a Rudolph Autopol I automatic polarimeter, and IR data were obtained using a Bruker Tensor 27 spectrometer. NMR data were obtained using a Bruker AM-500 spectrometer. HRESIMS and HPLC were performed on Agilent Q-TOF micro mass spectrometer and Agilent 1200 series, respectively. Macroporous resin D101 (Donghong Chemical Co., Ltd.) was used for column chromatography. TLC analysis was performed on silica gel plates (Marine Chemical Ltd.).

2.2. Plant Materials

The seeds of *C. tiglium* were collected in September 2020, from Guangxi Province, P.R. China, and were identified by Professor Jianyong Zhu. A voucher specimen (CT202009) was deposited at our laboratory.

2.3. Extraction and Isolation

The seeds of *C. tiglium* (2.5 kg) were pulverized and extracted with 90% ethanol (25 L), each 7 days at rt. The solvent was evaporated in vacuum to give an extract (395.0 g), which was suspended in water and separated with EtOAc. The EtOAc extract (312.0 g) was separated over macroporous resin column, eluting with MeOH-H₂O (5:5 → 9:1, v/v), to yield five fractions (Fr.A–Fr.E). Fr.C (42.2 g) was separated by CC on silica gel eluted with (petroleum ether-EtOAc, 15:1 → 1:2, v/v) afford Fr.C1–Fr.C5. Fr.C3 (1.2g) was separated over Sephadex column, and then purified via HPLC (MeOH-H₂O, 8:2, v/v) to give **3** (12.5 mg) and **4** (58.0 mg). Compound **1** (4.2 mg) and **2** (31.8 mg) were isolated from Fr.C4 (300.0 mg) in a similar way.

2.4. Spectral Data

Compound 1: colorless crystals, mp 166–168 °C; $[\alpha]_D^{25}$ -16.8 (*c* 0.25, MeCN); UV (MeCN) λ_{\max} (log ϵ) 223 (3.26), 195 (3.27) nm; IR (KBr) ν_{\max} 3405, 2927, 1693, 1376, 1261, 1248, 734 cm⁻¹; ¹H and ¹³C NMR data, see Table 1; HRESIMS *m/z* 411.1782 [M + Na]⁺ (calcd for C₂₂H₂₈O₆Na⁺, 411.1778).

2.5. X-ray Crystallographic Data of **1**

C₂₂H₂₈O₆ (*M* = 388.44 g/mol): orthorhombic, space group P2₁2₁2₁ (no. 19), *a* = 8.88430(10) Å, *b* = 9.46280(10) Å, *c* = 24.2964(3) Å, *V* = 2042.61(4) Å³, *Z* = 4, *T* = 100.00(10) K, μ (Cu K α) = 0.748 mm⁻¹, *D*_{calc} = 1.263 g/cm³, 20960 reflections measured (7.276° ≤ 2 θ ≤ 157.75°), 4323 unique (*R*_{int} = 0.0383, *R*_{sigma} = 0.0247) which were used in all calculations. The final *R*₁ was 0.0312 (*I* > 2 σ (*I*)) and *wR*₂ was 0.0824 (all data). Flack parameter = -0.01(6). The complete data were deposited at the Cambridge Crystallographic Data Centre (CCDC 2170690).

2.6. Cytotoxicity Assay

A375 human malignant melanoma cells were obtained from the cell bank of Chinese Academy of Sciences (Shanghai, China) and were cultivated in DMEM containing 10% FBS and 0.5% penicillin/streptomycin. The cells were incubated with 5% CO₂ at 37 °C in a humidified environment. The cytotoxicity for compounds **1–4** was measured by MTT method as our previously described [9].

3. Results and Discussion

3.1. Structure Elucidation

Compound **1** was obtained as colourless block crystals and assigned a molecular formula of C₂₂H₂₈O₆ by the HRESIMS positive peak at *m/z* 411.1782 [M + Na]⁺ (calcd for C₂₂H₂₈O₆Na⁺,

411.1778). The IR data of **1** showed absorptions at 3405 cm^{-1} (hydroxyl group) and 1693 cm^{-1} (carbonyl group). The ^1H NMR spectrum (Table 1) of **1** displayed one aldehyde group (δ_{H} 9.30, 1H, s, H-20); two olefinic protons at δ_{H} 7.01 (1H, br s, H-1) and 6.08 (1H, t, $J = 2.9$, H-7); one oxygenated methine (δ_{H} 4.05, 1H, d, $J = 9.7$, H-12); and five methyl groups at δ_{H} 1.25 (3H, s, Me-16), 1.29 (3H, s, Me-17), 1.29 (3H, d, $J = 6.3$ Me-18), 1.75 (3H, s, Me-19) and 2.10 (3H, s, Me-2'). The ^{13}C NMR spectrum of **1** showed 22 carbons comprising a ketocarbonyl group (δ_{C} 210.3), an aldehyde group (δ_{C} 194.2), a carbonyl group (δ_{C} 173.8), two double bonds (δ_{C} 155.0, 152.7, 143.6 and 140.3), a quaternary carbon, two oxygenated tertiary carbons, five methyls, six sp^3 methines (one oxygenated), and a sp^3 methylenes. The aforementioned spectroscopic data implied that compound **1** was a tigliane diterpenoid with an acetyl group [8].

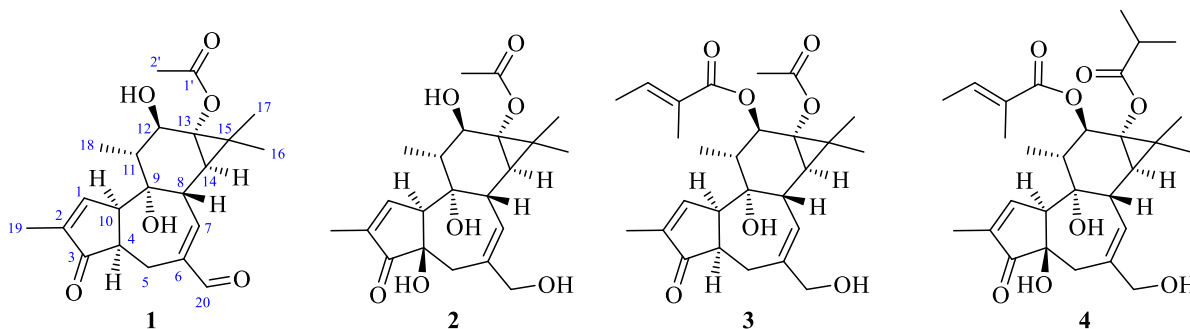


Figure 1. Structures of compounds **1–4** from *Croton tiglium*

In the ^1H - ^1H COSY spectrum, three fragments (C-1–C-10–C-4–C-5, C-7–C-8–C-14 and C-12–C-11–C-18) were established. The above fragments combined with HMBC correlations of Me-19/C-1, C-2 and C-3; H-1 and H-4/C-3; H-20/C-5, C-6 and C-7; H-7 and H-10/C-9; Me-18/C-9, C-11 and C-12; H-14/C-12 and C-13; Me-16 and Me-17/C-13, C-14 and C-15 indicated the existence of a typical 5/7/6/3-fused tigliane skeleton. The weak HMBC correlation from Me-2' to C-13 confirmed the location of the acetoxy group at C-13. Thus, the planar structure of **1** was defined as shown in Figure 2.

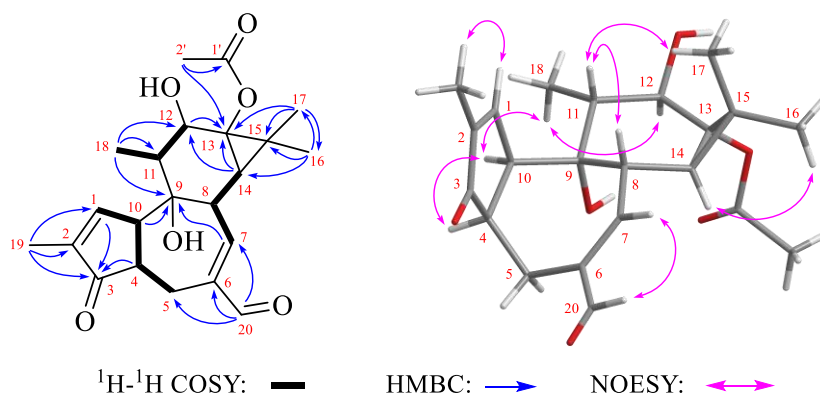


Figure 2. Key HMBC, ^1H - ^1H COSY and NOESY correlations for compound **1**

The relative configuration of **1** was assigned by the NOESY spectrum. The NOE interactions (Figure 2) of H-11/H-8 and Me-17 indicated that they were cofacial and defined to be β -orientation. Thus, H-4/H-10, H-10/Me-18, Me-18/H-12 and H-14/Me-16 revealed that H-4, H-10, H-12 and H-14 were α -orientations. In addition, the double bond between C-1 and C-2 was deduced as a *Z*-configuration by NOE interaction of H-1/Me-19, while the double bond between C-6 and C-7 was determined as *E*-configuration based on NOE correlation of H-7/H-20. Finally, it is absolute

configuration of 4*R*,8*S*,9*R*,10*R*,11*R*,12*R*,13*S*,14*R* (Figure 3) was confirmed by Cu K α single-crystal X-ray diffraction with the Flack parameter of $-0.01(6)$. Thus, the structure of **1** was assigned as 4-deoxy-20-oxophorbol-13-acetate.

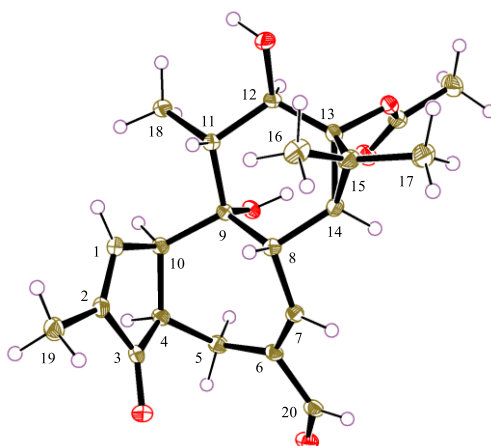


Figure 3. X-ray crystallographic structure for compound **1**

Table 1. ^1H (500 MHz) and ^{13}C NMR (125 MHz) data for compound **1** in CDCl_3

Position	δ_{H} (J in Hz; δ in ppm)	δ_{C}	Position	δ_{H} (J in Hz; δ in ppm)	δ_{C}
1	7.01, br s	155.0	12	4.05, d (9.7)	74.9
2		143.6	13		67.4
3		210.3	14	0.87, d (5.5)	34.9
4	2.84, m	48.4	15		26.0
5 α	3.05, ddd (15.9, 6.4, 2.9)	20.7	16	1.25, s	24.0
5 β	3.27, dd (15.9, 4.4)		17	1.29, s	16.4
6		140.3	18	1.29, d (6.3)	12.3
7	6.08, t (2.9)	152.7	19	1.75, s	10.5
8	2.25, m	41.6	20	9.30, s	194.2
9		77.7	1'		173.8
10	3.54, m	47.1	2'	2.10, s	21.1
11	1.64, m	44.9			

The known compounds, phorbol-13-acetate (**2**) [10], 12-*O*-tiglyl-4 α -deoxyphorbol-13-acetate (**3**) [11] and 12-*O*-tiglylphorbol-13-isobutyrate (**4**) [12] were identified via comparing their data with literature values.

3.2. Cytotoxic Activity

All compounds (**1–4**) were screened for cytotoxicities against A375 cells using the MTT assay, and dacarbazine was used as a positive drug ($\text{IC}_{50} = 23.5 \pm 2.8 \mu\text{M}$). The results displayed that compounds **1**, **3** and **4** exhibited moderate cytotoxic activities with IC_{50} values of 32.6 ± 3.5 , $18.5 \pm 1.8 \mu\text{M}$ and $26.4 \pm 2.2 \mu\text{M}$, respectively, while **2** had no obvious cytotoxicity ($\text{IC}_{50} > 50 \mu\text{M}$).

Acknowledgments

This work was supported by the Anhui Province Education Department Nature and Science Fund Key Project of China, Hefei, China (No. KJ2018ZD065) and the Special Fund for Modern Chinese

medicine technology innovation and social service team of Bozhou Vocational and Technical College (Nos. yptd001 and ypz002).

Supporting Information

Supporting information accompanies this paper on <http://www.acgpubs.org/journal/records-of-natural-products>

ORCID

Lijuan Zhang: [0000-0003-2840-8668](https://orcid.org/0000-0003-2840-8668)

Fei Li: [0000-0001-5737-8555](https://orcid.org/0000-0001-5737-8555)

Jianyong Zhu: [0000-0001-5922-9326](https://orcid.org/0000-0001-5922-9326)

Qian Niu: [0000-0003-2557-9285](https://orcid.org/0000-0003-2557-9285)

References

- [1] H. B. Wang, X. Y. Wang, L. P. Liu, G. W. Qin and T. G. Kang (2015). Tiglane diterpenoids from the *Euphorbiaceae* and *Thymelaeaceae* families, *Chem. Rev.* **115**, 2975–3011.
- [2] Editor Committee for Flora of China of Chinese Academy of Science (1996). Science Publishing House, Beijing. pp. 133.
- [3] J. F. Wang, S. H. Yang, Y. Q. Liu, D. X. Li, W. J. He, X. X. Zhang, Y. H. Liu and X. J. Zhou (2015). Five new phorbol esters with cytotoxic and selective anti-inflammatory activities from *Croton tiglium*, *Bioorg. Med. Chem. Lett.* **25**, 1986–1989.
- [4] B. Q. Zhao, S. Peng, W. J. He, Y. H. Liu, J. F. Wang and X. J. Zhou (2016). Antitubercular and cytotoxic tiglane-type diterpenoids from *Croton tiglium*, *Bioorg. Med. Chem. Lett.* **26**, 4996–4999.
- [5] S. El-Mekkawy, M. R. Meselhy, N. Nakamura, M. Hattori, T. Kawahata and T. Otake (2000). Anti-HIV-1 phorbol esters from the seeds of *Croton tiglium*, **53**, 457–464.
- [6] J. Wang, L. Qin, B. Zhao, L. Cai, Z. Zhong, Y. Liu and X. Zhou (2019). Crotonols A and B, two rare tiglane diterpenoid derivatives against K562 cells from *Croton tiglium*, *Org. Biomol. Chem.* **17**, 195–202.
- [7] X. L. Zhang, L. Wang, F. Li, K. Yu and M. K. Wang (2013). Cytotoxic phorbol esters of *Croton tiglium*, *J. Nat. Prod.* **76**, 858–864.
- [8] Q. Du, Y. Zhao, H. Liu, C. Tang, M. Zhang, C. Ke and Y. Ye (2017). Isolation and structure characterization of cytotoxic phorbol esters from the seeds of *Croton tiglium*, *Planta Med.* **83**, 1361–1367.
- [9] G. K. Wang, Y. P. Sun, W. F. Jin, Y. Yu, J. Y. Zhu and J. S. Liu (2022). Limonoids from *Swietenia macrophylla* and their antitumor activities in A375 human malignant melanoma cells, *Bioorg. Chem.* **123**, 105780.
- [10] D. S. Yang, Q. X. He, Y. P. Yang, K. C. Liu and X. L. Li (2014). Chemical constituents of *Euphorbia tibetica* and their biological activities, *Chin. J. Nat. Med.* **12**, 38–42.
- [11] W. Dou, Y. Hao, J. Liu, D. Yuan and H. Fu (2016). Two novel phorbol esters from *Croton tiglium* L., *J. Chin. Pharm. Sci.* **25**, 771–778.
- [12] G. T. Marshall and A. D. Kinghorn (1984). Short-chain phorbol ester constituents of croton oil, *J. Am. Oil Chem. Soc.* **61**, 1220–1225.

A C G
publications

© 2022ACG Publications