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# **Chemical Constituents from the Whole Plant**

## of Pachysandra terminalis

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Abstract: Two new compounds, butyl(Z)-3-((3R,4R)-5-(hydroxymethyl)-3,4-dimethyl-3,4-dihydro-2*H*-pyran-6-yl)acrylate (1) and (2Z,4S)-4,5-dihydroxy-3,4-dimethylpent-2-enoic acid (2) along with seven known ones, stigmast-5,28(29)-dien-3 $\beta$ -ol (3),  $\beta$ -sitosterol (4), carotene (5), fraxetin (6), *p*-coumaric acid (7), cis-*p*-hydroxycinnamic acid (8), ferulic acid (9) were obtained from the whole plant of *Pachysandra terminalis*. The structures of these compounds were elucidated by comprehensive spectroscopic methods including 1D, 2D NMR, MS, IR and ECD data analysis. Notably, compounds 3 and 5~8 were isolated from genus *Pachysandra* for the first time. Moreover, compounds 1~3 and 6~8 were tested their cytotoxic activities against three cancer cells, however, only compound 1 showed inhibitory effect in SW620 cells with IC<sub>50</sub> value of 47.7  $\mu$ M.

**Keywords**: *Pachysandra terminalis*; chemical composition; isolation and purification; steroids; fatty acids. © 2022 ACG Publications. All rights reserved.

## 1. Introduction

Pachysandra terminalis, an evergreen plant, belongs to the genus Pachysandra, family Buxaceae [1-2], which is widely distributed in the South of China. It is mainly distributed in the Qinba Mountains, Shaanxi province of China [3]. The chemical constituents isolated from P. terminalis are mainly Pachysandra-type alkaloids, triterpenoids, volatile oils, and others [4]. Modern pharmacological studies have shown that it has antioxidant, anti-ulcer [5-7], anti-tumor [8-13], anti-bacterial [14-15], and insecticidal [16-17] activities. It was mainly used for the clinical treatment of rheumatoid arthritis and chronic bronchitis. In our continuous work to investigate more bioactive natural compounds from Р. terminalis, two new compounds butyl(Z)-3-((3R,4R)-5-(hydroxymethyl)-3,4-dimethyl-3,4-dihydro-2H-pyran- 6-yl)acrylate (1) and (2Z,4S)-4,5-dihydroxy-3,4-dimethylpent-2-enoic acid (2) and seven known compounds, stigmast-5,28(29)-dien- $3\beta$ -ol (3) [18],  $\beta$ -sitosterol (4) [19], carotene (5) [20], fraxetin (6) [21],

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*p*-coumaric acid (7) [22], *cis-p*-hydroxycinnamic acid (8) [23], ferulic acid (9) [24], were procured (Figure 1). In this study, we described the structure identification and cytotoxic activities of these compounds.



Figure 1. Structures of compounds 1–9

## 2. Materials and Methods

## 2.1. General Experimental Procedures

The HR-ESI-MS spectra was taken on an Agilent Technologies 6550 Q-TOF and ESI-MS was performed on Waters Quattro Premier instrument. 1D and 2D NMR spectra were recorded on a Bruker-AVANCE 400 instrument with TMS as an internal standard. Semipreparative HPLC was performed on a system comprising an LC-20AP pump equipped with a SPD-20A UV detector and a Ultimate XB-C<sub>18</sub> (10 mm × 250 mm, 5  $\mu$ m particles). Sephadex LH-20 gel silica gel were purchased from GE Healthcare Bio-Sciences AB. Chromatographic methanol (Tianjin Comio Chemical Reagent Co., Ltd.)

## 2.2. Plant Material

In the present study, *Pachysandra terminalis* Sieb. et Zucc. were collected from the Baoji, Shaanxi Province, China, in 2020, and authenticated by Professor Wei Wang (School of Pharmacy, Shaanxi University of Chinese Medicine). A voucher specimen (herbarium No. 20200901) has been deposited in the Medicinal Plants Herbarium, Shaanxi University of Chinese Medicine, Xianyang, China.

#### 2.3. Extraction and Isolation

The whole plant of *P. terminalis* (10.0 kg) was extracted with 80 % EtOH under reflux three times. After removal of EtOH solvent under reduced pressure, the extract was suspended in water and successively extracted with petroleum ether,  $CH_2Cl_2$  and n-BuOH. The  $CH_2Cl_2$  parts (160 g) were chromatographed on silica gel column, eluted with gradient solvent system ( $CH_2Cl_2$ - $CH_3OH$ , 80:1–0:1) to give thirteen fractions (Fr.1-Fr.13).

Fr. 11 (30.0 g) was subjected to Sephadex LH-20 column chromatography and eluted with CH<sub>2</sub>Cl<sub>2</sub>-CH<sub>3</sub>OH (1:1) to yield Fr. 11-1~ Fr. 11-5, Fr. 11-1 (3.5 g) was purified by SP-HPLC with CH<sub>3</sub>OH-H<sub>2</sub>O (20:80) as mobile phase to obtained compounds **2** ( $t_R$  = 18.0 min, 16.5 mg), **9** ( $t_R$  = 25.3 min, 20.0 mg), **7** ( $t_R$  = 35.0 min, 14.0 mg) and **8** ( $t_R$  = 35.5 min, 17.0 mg). Fr. 11-3 (1.8 g) was purified by SP-HPLC with CH<sub>3</sub>OH-H<sub>2</sub>O (25:75) as mobile phase to obtained compounds **3** ( $t_R$  = 35.0 min, 21.6 mg) and **4** ( $t_R$  =4 0.5 min, 25.0 mg). Fr. 11-4 (3.0 g) was purified by SP-HPLC with CH<sub>3</sub>OH-H<sub>2</sub>O (25:75) as mobile phase to obtained compound **6** ( $t_R$  = 30.5 min, 17.0 mg). Fr. 11-5 (0.5 min, 21.0 mg).

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g) was purified by SP-HPLC with CH<sub>3</sub>OH-H<sub>2</sub>O (20:80) as mobile phase to obtained compounds 5 ( $t_R$  = 30.0 min, 30.6 mg) and 1 ( $t_R$  = 40.5 min, 7.0 mg) (Figure 1).

## 2.4. Spectroscopic Data

Butyl(Z)-3-((3R,4R)-5-(hydroxymethyl)-3,4-dimethyl-3,4-dihydro-2H-pyran-6-yl)acrylate (1): A reddish brown oily solid,  $[\alpha]_D^{20}$ -11.2 (*c* 0.05, CH<sub>3</sub>OH); IR v<sub>max</sub> (KBr) (cm<sup>-1</sup>): 3305, 2950, 2834, 1735 and 1452 cm<sup>-1</sup>; <sup>1</sup>H-NMR (400 MHz, CH<sub>3</sub>OH) and <sup>13</sup>C-NMR (100 MHz, CH<sub>3</sub>OH) spectral data, see Table 1; HR-ESI-MS: *m/z* 269.1748 [M+H] <sup>+</sup> (calcd. for C<sub>15</sub>H<sub>25</sub>O<sub>4</sub>, 269.1753).

(2Z, 4S)-4,5-dihydroxy-3,4-dimethylpent-2-enoic acid (2): A yellow oily solid,  $[\alpha]_D^{20}$  -2.8 (c 0.05, CH<sub>3</sub>OH); IR  $\nu_{max}$  (KBr) (cm<sup>-1</sup>): 3342, 2938, 2883, 1738, 1430 and 1032; <sup>1</sup>H-NMR (400 MHz, CH<sub>3</sub>OH) and <sup>13</sup>C-NMR (100 MHz, CH<sub>3</sub>OH) spectral data, see Table 2; HR-ESI-MS: *m*/z 161.0814 [M+H] <sup>+</sup> (calcd. for C<sub>7</sub>H<sub>13</sub>O<sub>4</sub>, 161.0814).

## 3. Results and Discussion

## 3.1. Structure Elucidation

Compound 1 was isolated as a reddish brown oily solid. The molecular formula  $C_{15}H_{25}O_4$  was supported by the positive HR-ESI-MS molecular ion peak at m/z 269.1748 [M+H]<sup>+</sup> (calculated 269.1753 [M+H]<sup>+</sup>). The IR spectrum displayed for hydroxy (3305 cm<sup>-1</sup>), carbonyl (1735 cm<sup>-1</sup>) and double bonds (1452 cm<sup>-1</sup>). The <sup>1</sup>H NMR data of 1 (Table 1) exhibited three methyl signals at  $\delta_H$  0.98 (3H, t, CH<sub>3</sub>-4'), 1.00 (3H, d, J = 7.8 Hz, CH<sub>3</sub>-12), 1.06 (3H, d, J = 7.8 Hz, CH<sub>3</sub>-11), three hypoxia-methylene signals at  $\delta_H$  3.58, 3.88 (2H, d, J = 2.5 Hz, H-10), 3.33 and 3.99 (2H, t, J = 8.2Hz, H-2), 4.29 (2H, t, J = 6.6 Hz, H-1') and a double bond signal at  $\delta_H$  7.62 (1H, d, J = 10.1 Hz, H-7) and 7.72 (1H, d, J = 10.1 Hz, H-8). The <sup>13</sup>C NMR data of 1 (Table 1) displayed 15 carbon signals, three of which belongs to the methyl groups at ( $\delta_C$  11.8, 14.1, 15.9), three of which were determined as methylene groups at ( $\delta_C$  64.5, 66.7, 74.4), four of which were confirmed as two double bond signals at ( $\delta_C$  106, 133.6, 129.9, 132.4) and carbonyl carbon signal at  $\delta_C$  169.3. In addition, five of which were determined as methylene groups at ( $\delta_C$  20.3, 31.7, 64.5, 66.7, 74.4) in the DEPT-135 spectrum. The 2D NMR data analysis confirmed the conclusion above.

The <sup>1</sup>H-<sup>1</sup>H COSY correlations (Figure 2) from H-12/H-3, H-3/H-4 and H-4/H-11, accompanied with the HMBC correlations (Figure 2) of H-12/C-2, C-3 and C-4, H-11/C-3, C-4 and C-5, H-10/C-4, C-5 and C-6, the six-membered ring is an alkene ether structure based on C-6 ( $\delta_{\rm C}$ 133.6) and C-2 ( $\delta_{\rm C}$  74.4), C-5 ( $\delta_{\rm C}$  106.8) is connected with hydroxymethyl. In the HMBC spectrum (Figure 2), the correlation between the proton signal at H-7 ( $\delta_{\rm H}$  7.62) with the carbon signal at C-6 ( $\delta_{\rm C}$ 133.6), suggested that two double bonds are connected through C-6 and C-7. In the <sup>1</sup>H-<sup>1</sup>H COSY spectrum (Figure 2), there is a correlation between H-7 and H-8, accompanied with the HMBC correlations (Figure 2) of H-8/C-9 and C-7, description of the double bond and the C-9 ( $\delta_{\rm C}$  169.3) carbonyl group related. Finally, H-4'/H-3', H-2' and H-1' was found in the <sup>1</sup>H-<sup>1</sup>H COSY correlation spectrum, and the HMBC spectrum (Figure 2) shows that H-1'/C-9, C-2' and C-3', indicates that the carbonyl group (C-9) is linked to the n-butanol group, which is the n-butanol ester, demonstrated the 2D structure of 1 as butyl-3-(5-(hydroxymethyl)-3,4-dimethyl-3,4-dihydro-2H-pyran-6-yl) acrylate. In the NOESY spectrum of 1 (Figure 2), correlations between H-11/H-3 and H-12/H-4 deduced the  $\beta$ -configuration of H-3 and CH<sub>3</sub>-11 and  $\alpha$ -configuration of CH<sub>3</sub>-12 and H-4. Coupling constants of  $J_{7,8} = 10.1$  Hz confirmed the Z configuration of  $\Delta^{7,8}$ . To further determine the absolute configuration of 1, the ECD curves (Figure 3) were simulated of 1 [(3R, 4R)-1 and (3S, 4S)-1]. The experimental and calculated ECD curves of (3R, 4R)-1 matched well [25]. Thus, the structure of 1 was assigned as butyl(Z)-3-((3R,4R)-5-(hydroxymethyl)- 3,4-dimethyl-3,4-dihydro-2H-pyran-6-yl) acrylate (Figure 1).

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FOSITION OC	$\delta_{ m H}$	[	Position	$\delta_{\mathrm{C}}$	$\delta_{ m H}$
2 74.4	4, $CH_2$ 2a 2b	, 3.33, t, (8.2) , 3.99, t, (8.2)	10	64.5, CH <sub>2</sub>	3.88, d, (2.5)
<b>3</b> 52.0	0, CH 1.4	46, m	11	11.8, CH <sub>3</sub>	1.06, d, (7.8)
4 40.4	4, CH 2.1	11, m	12	15.9, CH <sub>3</sub>	1.00, d, (7.8)
5 106	5.8, C —	-	1′	66.7, CH <sub>2</sub>	4.29, t, (6.6)
<b>6</b> 133	8.6, C —	-	2'	31.7, CH <sub>2</sub>	1.72, m
7 132	2.4, CH 7.0	62, d, (10.1)	3'	$20.3, CH_2$	1.44, m
8 129	9.9, CH 7.'	72, d, (10.1)	4'	14.1, CH <sub>3</sub>	0.98, t, (7.5)
<b>9</b> 169	0.3, C —	-			

Table 1. <sup>1</sup>H-NMR (400 MHz, in CD<sub>3</sub>OD) and <sup>13</sup>C-NMR (100 MHz, in CD<sub>3</sub>OD) spectral data of compound 1



Figure 2. Key <sup>1</sup>H - <sup>1</sup>H COSY, HMBC and NOESY correlations of compound 1



Figure 3. Experimental and calculated ECD spectra of 1

Compound **2** was isolated as a yellow oily solid. The molecular formula  $C_7H_{13}O_4$  was supported by the positive HR-ESI-MS molecular ion peak at m/z 161.0814 [M+H]<sup>+</sup> (calculated 161.0814 [M+H]<sup>+</sup>). The IR spectrum displayed for hydroxy (3342 cm<sup>-1</sup>), carbonyl (1738 cm<sup>-1</sup>) and

double bonds (1430 cm<sup>-1</sup>). The <sup>1</sup>H NMR data of **2** (Tab. 2) exhibited two methyl signals at  $\delta_{\rm H}$  1.38 (3H, s, CH<sub>3</sub>-4), 2.07 (3H, d, J = 1.2 Hz, CH<sub>3</sub>-3), one hypoxia-methylene signals at  $\delta_{\rm H}$  3.70 (2H, m, H-5) and a trisubstituted double bond at  $\delta_{\rm H}$  5.83 (1H, d, J = 1.4 Hz, H-2). The <sup>13</sup>C NMR data of **2** (Table 2) displayed 7 carbon signals, two of which belongs to the methyl groups at ( $\delta_{\rm C}$  11.8, 18.2), one of which were determined as methylene groups at  $\delta_{\rm C}$  63.9, two of which were confirmed as double bond signals at ( $\delta_{\rm C}$  116.7, 172.0) and carbonyl carbon signal at  $\delta_{\rm C}$  173.6. In addition, one of which were determined as methylene groups at  $\delta_{\rm C}$  63.9 in the DEPT-135 spectrum. The 2D NMR data analysis confirmed the conclusion above. The HMBC correlations (Figure 4) correlations from CH<sub>3</sub>-3/C-2, C-3 and C-4, prove the correlation between double bond and methyl group, CH<sub>3</sub>-4/C-3, C-4 and C-5, H-5/C-3, C-4 and CH<sub>3</sub>-4, H-2/CH<sub>3</sub>-3, C-4 and C-1 disclosed the 2D structure of **2** as 4,5-dihydroxy-3,4-dimethylpent- 2-enoic acid. In the NOESY spectrum of **2**, correlations between H-2 and H-3(CH<sub>3</sub>-3), indicated the *Z* configuration of  $\Delta^{2,3}$ . To further determine the absolute configuration of **2**, the ECD curves (Figure 5) were simulated of **2** [(4*S*)-**2** and (4*R*)-**2**]. The experimental and calculated ECD curves of (4*S*)-**2** matched well [26]. Therefore, the structure of **2** was assigned as (2*Z*,4*S*)-4,5- dihydroxy-3,4-dimethylpent-2-enoic acid (Figure 1).

Table 2. <sup>1</sup>H-NMR (400 MHz, in CD<sub>3</sub>OD) and <sup>13</sup>C-NMR (100 MHz, in CD<sub>3</sub>OD) spectral data of compound 2

Position	$\delta_{ m C}$	$\delta_{ m H}$	
1	173.6, C		
2	116.7, CH	5.83, d, (1.4)	
3	172.0, C		
4	90.7, C		
5	63.9, CH <sub>2</sub>	3.70, m	
CH <sub>3</sub> -3	11.8, CH <sub>3</sub>	2.07, d, (1.2)	
CH <sub>3</sub> -4	18.2, CH <sub>3</sub>	1.38, s	



Figure 4. Key HMBC and NOESY correlations of compound 2

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Figure 5. Experimental and calculated ECD spectra of 2

### 3.2. Cytotoxicity Assay

The cytotoxic activities assay toward three human tumor cell (A549, HCT116 and SW620) lines were measured by the MTT method as we reported previously [4] for compounds  $1\sim3$  and  $6\sim8$ , using cisplatin as positive control. The experimental results (Tab. 3) showed that these compounds showed weak cytotoxicity in the human cancer cell lines.

Compounds	A549	HCT116	SW620
Cisplatin	$32.1\pm1.3$	$43.5\pm3.3$	$32.5 \pm 3.4$
1	>100	>100	$47.7 \pm 2.5$
2	>100	>100	>100
3	>100	>100	>100
6	>100	>100	>100
7	>100	>100	>100
8	>100	>100	>100

**Table 3.** Cytotoxic activities of compounds  $1 \sim 3$  and  $6 \sim 8$  on A549, HCT116 and SW620 cancer cell lines. (IC<sub>50</sub>,  $\mu$ M)<sup>a</sup>

 ${}^{a}IC_{50}$  values are means from three independent experiments (average  $\pm$  SD) in which each compound concentration was tested in three replicate wells; Cisplatin as positive control.

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### **Supporting Information**

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